

# Guayule (*Parthenium argentatum*) resin: A review of chemistry, extraction techniques, and applications

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## ABSTRACT

Guayule (*Parthenium argentatum* Gray) is a perennial shrub of the Asteraceae family native to the arid southwestern U.S. and northern Mexico that produces high-quality, hypoallergenic natural rubber. Guayule processing produces three output streams: high-molecular weight rubber as the primary product, a mixture of resin/low-molecular weight rubber, and a fine, dry woody bagasse. The goal of this review was to evaluate guayule resin as a source of natural resin and secondary metabolites in order to overcome obstacles to commercialization by maximizing the total economic value of extractable fractions, increasing the scale and robustness of extraction methods, while concurrently lowering the operational costs and environmental impacts of separations. Based on the literature and current market prices, the guayule resin applications with the highest economic potential are residential pesticides (with a specific price range of 1.1–6.8 USD/kg), paints and coatings (0.7–6.0 USD/kg), wood preservative coatings (0.8–6.0 USD/kg), amine-epoxy strippable coatings (0.8–2.5 USD/kg), and adhesives (0.125–1.60 USD/kg). The separation methods with the greatest potential for scale-up are pressurized solvent extractions, such as supercritical CO<sub>2</sub> extraction, due to their reduced extraction times, energy requirements, and solvent consumption.

## 1. Introduction

Guayule is a perennial shrub of the Asteraceae family (over 20,000 species) native to the arid region of the southwestern U.S. and north-central Mexico. Guayule is a source of high-quality natural rubber (*cis*-1,4-polyisoprene), mostly identical to the rubber from the *Hevea brasiliensis* tree, which supplies 90 % of the world's natural rubber for applications in the tire, medical, pharmaceutical, and food industries (Mooibroek and Cornish, 2000). Annual global consumption of natural rubber is more than 12 megatons; by 2030, consumption is predicted to rise to 30 megatons (Cornish, 2017). Natural latex rubber from *Hevea* trees contains the protein responsible for type 1 latex allergies (Ownby et al., 1996). Guayule rubber does not contain the allergenic proteins, providing the potential for hypoallergenic latex materials (Cornish, 1996). Guayule also does not contain laticifers as *H. brasiliensis* does; the rubber is stored in the cytosol of parenchyma tissues (Backhaus and Walsh, 1983) and mesophyll cells of leaves (Madhavan and Benedict, 1984), and thus requires different methods for rubber extraction. An

acre of two-year-old guayule plants will typically yield 0.5–1 Mg of rubber (Cornish et al., 2006). Guayule plants can tolerate some intense growing conditions: temperatures ranging from −18 °C to +49 °C (van Beilen and Poirier, 2007), rainfall as low as 350–800 mm/year, a variety of soils, and pest pressures (Foster and Coffelt, 2005).

Despite all of these advantages, commercialization of guayule rubber has been slow. Among the reasons for poor commercialization of guayule are the low cost of *Hevea* rubber (approximately 1.30–2.50 USD/kg in 2020); low rubber yield (around 5–7 % by weight for a 3- to 5-year-old shrub); significant costs of cultivation, harvesting, and processing; and no efficient value-added use of the co-products (resin and bagasse). According to a recent techno-economic analysis of guayule rubber production (Sproul et al., 2020), a baseline minimum selling price of 3.08 USD/kg of guayule rubber was reported, with resin and bagasse being sold at 1.00 USD/kg and 0.10 USD/kg, respectively. The three parameters that largely impact rubber production cost are harvest yield, biomass rubber content, and resin value; improving the resin value can reduce the minimum selling price of guayule rubber as much as 50 %

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(Sproul et al., 2020). The feasibility of guayule in the Mediterranean region was investigated by Sfeir et al. (2014) and Snoeck et al. (2015).

The first record of guayule rubber use was in the early 1500s (pre-European contact) by native peoples of Mexico for making balls for use in games (Lloyd, 1911). Guayule was first used for commercial production of rubber in the late 1800s (Bonner, 1991; Singh, 2010). Since then, there have been four general periods of guayule commercialization efforts in North America, summarized in Fig. 1.

Guayule rubber can be extracted from the dried shrub using solvent extraction, or from freshly-harvested shrub using aqueous (latex) extraction (Cornish, 1996; Cornish and Backhaus, 1990; Jones, 1948; Schloman et al., 1996). A resinous liquid (a mixture of low-molecular-weight rubber and secondary metabolites) and bagasse are the residues from guayule rubber extraction (Wagner and Schloman, 1991). The physical properties of natural rubber are strongly influenced by the non-rubber constituents of the guayule plant (Monadjemi et al., 2016). As the non-rubber components, especially lipids (fatty acids like linoleic and linolenic, and triglycerides), facilitate the degradation of rubber (mostly by oxidation of polymer chain) (Keller et al., 1981), the separation of resin is a crucial step for producing high-quality rubber from guayule. A high content of resin in extracted dry rubber lowers the bulk viscosity of the final rubber product, so only a selective extraction and deseresination process results in a rubber product similar to *Hevea* rubber (Schloman, 2005).

A substantial amount of resin (generally more than the amount of rubber) is biosynthesized in guayule (Abdel-Haleem et al., 2018), about 6–15% of the dry weight of the plant (Schloman, 1988a, 1988b). Both rubber and resin are produced in the epithelial cells of resin ducts (Joseph et al., 1988). Resin is accumulated in schizogenous canals and secreted into the duct lumen in leaves, stems, and roots (Gilliland et al., 1988; Joseph et al., 1988). Resin yield from acetone extraction varies significantly from 5.5–11 wt.% (Sidhu et al., 1995) based on cultivation site, harvest date, shrub strain, shrub age, and processing procedures (Schloman et al., 1986). Resin concentration has been reported at 5.5–7 dry wt.% for 2-year-old plants, and 6–8.5 dry wt.% for 3- and 4-year-old plants, but there is no correlation between total biomass yield and resin concentration (Estilai, 1991; Sidhu et al., 1995).

Unlike other natural resins, which are extracted as the main product and are the primary source of revenue (e.g. balsam and pine), guayule resin currently contributes little to the economic feasibility of guayule as an industrial crop. The objective of this review is to define efficient separation and characterization methods for natural resin compounds to guide the development of applications for guayule resin at commercial

scale. A secondary objective of this review is to evaluate the economic and environmental case for large-scale natural rubber and resin extraction.

## 2. Guayule resin composition

Guayule resin contains a wide variety of secondary metabolites (sesquiterpene esters, triterpene alcohols, fatty acids) and is soluble in polar solvents such as alcohols, esters, and ketones. Guayule resin composition varies with shrub strain, harvest date, cultivation site, and processing history (Schloman et al., 1986). Among the limited studies on guayule resin, inconsistency between results has been linked to differences in extraction and pretreatment methods, on top of factors like genotype, plant age at the time of harvest, and cultural practices (Cheng et al., 2020). These factors have been shown to affect levels of sesquiterpene esters and fatty acid triglycerides (Schloman et al., 1983). Around 50 different classes of chemicals have been identified in guayule resin (Banigan et al., 1982). These are often divided into three main groups: terpenes/terpenoids, fatty acids, and low-molecular-weight rubber (LMWR).

### 2.1. Essential oils

The major groups of volatile fractions in the essential oil extracted from fresh guayule biomass include terpenes (70–83 %), oxygenated terpenes (6–10 %), sesquiterpenes (8–14 %), and sesquiterpene alcohols (3–6 %);  $\alpha$ -pinene is the most dominant compound (Haagen-Smit and Siu, 1944; Scora and Kumamoto, 1979). These volatile fractions are produced more rapidly than rubber during periods of guayule vegetative growth (Schloman et al., 1986). These volatile compounds are synthesized via methylerythritol phosphate (MEP) and mevalonic acid pathways (Fig. 2). Geranyl diphosphate (GPP), the precursor for all monoterpenes ( $C_{10}$ ), comes from the combination of dimethylallyl diphosphate (DMAPP) and isopentenyl diphosphate (IPP).

### 2.2. Sesquiterpene esters

The major sesquiterpene compounds, representing 10–15 wt.% of the resin, are guayulins (Sidhu et al., 1995; Watkins et al., 1985). Guayulins are both isoprenoids and aromatic acid esters (Teetor et al., 2009). Guayulins A and B are sesquiterpene esters of *trans*-cinnamic and *p*-anisic acids, respectively. A whole guayule resin contains approximately 8–14 wt.% of guayulin A and 0.5–3 wt.% of guayulin B, mostly concentrated in the woody stems (Schloman et al., 1983; Sidhu et al.,

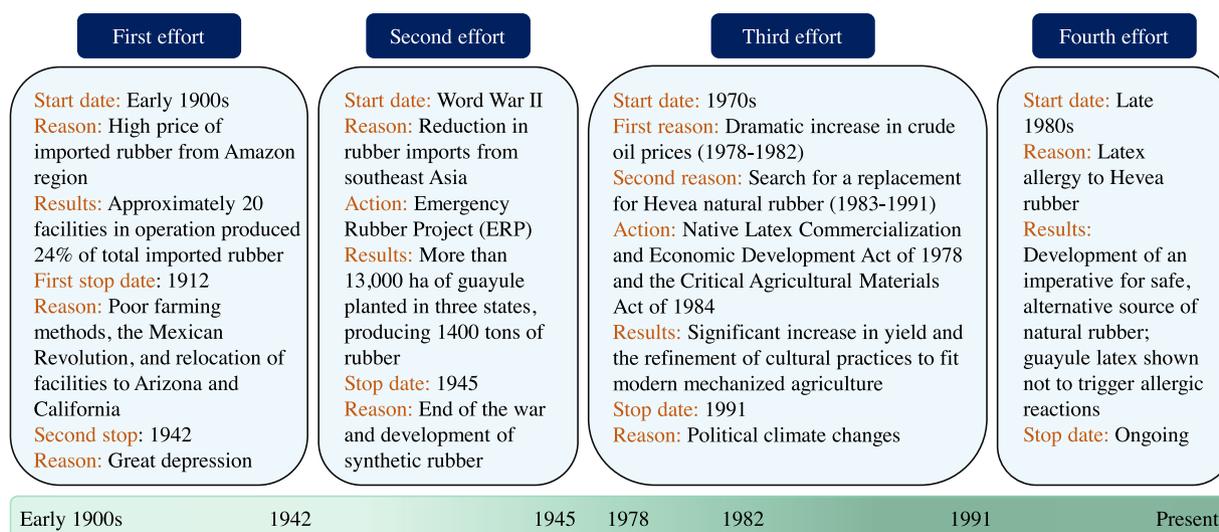


Fig. 1. Four periods of commercialization effort for guayule rubber. Elaborated from Bonner (1991); Huang (1991); Wright et al. (1991); Ray (1993); Foster and Coffelt (2005); Ownby et al. (1996); Siler et al. (1996); Cole et al. (1991); Finlay (2009), and Jones (1948).

1995; Teetor et al., 2009). The concentration of guayulin A is highly dependent on season (Schloman et al., 1986). Guayulins C and D are likely formed spontaneously through oxidation/degradation pathways of guayulins A and B, and constitute 0.1–3 wt.% and 0.1–1 wt.% of the resin, respectively (Rozalén et al., 2021; Schloman et al., 1983). The highest concentrations of guayulin C (1400–1650 mg/kg) and guayulin D (330–370 mg/kg) are found in the leaves (Spano et al., 2018). Rubber and guayulin A contents in stems with diameters larger than 10 mm are significantly correlated, so guayule A content could serve as a good predictor of total plant rubber content (Teetor et al., 2009).

Farnesyl diphosphate (FPP), the precursor of sesquiterpenes (C<sub>15</sub>), comes from addition of two IPP molecules to DMAPP (Fig. 2). Sesquiterpenes are found naturally as hydrocarbons, or in oxygenated forms, including lactones, alcohols, acids, aldehydes, and ketones (Awouafack et al., 2013).

### 2.3. Triterpenoid esters

Argentatins, tetracyclic-type triterpenes, are also abundant in guayule resin, constituting 20–30 % of the total resin (Martínez et al., 1990; West et al., 1991). Argentatins A–D and iso-agentatin B are the most dominant compounds in this class. These triterpenes have shown anti-inflammatory and cytotoxic activities (Alcántara-Flores et al., 2015; Tavares-Santamaría et al., 2020). Triterpenes (C<sub>30</sub>) are derived from dimerization of FPP via mevalonic acid pathways (Fig. 2).

### 2.4. Fatty acids

Fatty acid triglycerides encompass 10–25 wt.% of guayule resin (Banigan and Meeks, 1953; Keller et al., 1981). Linoleic acid (C<sub>18:3</sub>) is the most abundant at 36–65 % of the total fatty acids, distantly followed by palmitic, linolenic, oleic, and stearic acids. Cinnamic acid and *p*-anisic acid were identified as two major aromatic acids in the saponifiable materials (Schloman et al., 1983). Chemical treatments before and during extraction, such as sodium hydroxide in latex extraction, can significantly change (mostly decrease) the yield of fatty acids extracted from resin (Belmares et al., 1980). Fatty acids ranging from 8 to 32 carbons have been found in some plants, synthesized by carboxylation of acetyl-CoA to form malonyl-CoA under a repeated series of reactions.

### 2.5. Sterols

Sterols (e.g.  $\beta$ -sitosterol) are another major compound class in guayule resin (Cheng et al., 2020), accounting for 9–11 wt.% (Buchanan et al., 1978; Schloman et al., 1983). Plant sterols are synthesized from 2,3-oxidosqualene (C<sub>30</sub>H<sub>50</sub>O) via the MVA pathways (Fig. 2) from the cycloartenol (C<sub>30</sub>H<sub>50</sub>O) precursors. Plant sterols are formed by a tetracyclic ring and a long chain attached at C-17.

### 2.6. Low-molecular-weight rubber (LMWR)

LMWR is an integral part of guayule resin. Approximately 20–40 wt.% of low-molecular-weight *cis*-1,4-polyisoprene compounds can be found in the more polar (resin) phase after extraction and precipitation of high-molecular-weight rubber (Wagner and Schloman, 1991). LMWR has a molecular weight one to two orders of magnitude smaller than those of natural rubber polymers ( $1 \times 10^5$ – $2.5 \times 10^6$  g/mol) (Angulo-Sánchez et al., 1995; Campos-López and Angulo-Sánchez, 1976). Detection and characterization of LMWR compounds is difficult, even with advanced mass spectroscopy techniques, such as high-resolution Fourier transform ion cyclotron resonance mass spectroscopy (FT-ICR MS), which can provide accurate masses of hundreds of compounds with a wide range of molecular weights and degrees of aromaticity (Cheng et al., 2020).

### 2.7. Water extractables

Resin may contain (depending on the extraction method) water extractables, which constitute 14–17 % of the woody tissues (Buchanan et al., 1978; Schloman et al., 1983; Traub and Slattery, 1946). On a whole-shrub basis, bagasse yielded 16 % extractables (7.5 % polyphenolics and 8.5 % polysaccharides) after sequential extraction, and 19 % extractables (6.8 % polyphenolics and 12.6 % polysaccharides) after simultaneous extraction (Schloman et al., 1988). The substantially lower level of polysaccharides in bagasse after sequential extraction results from entrainment of polysaccharides into the resin as part of the free aqueous phase.

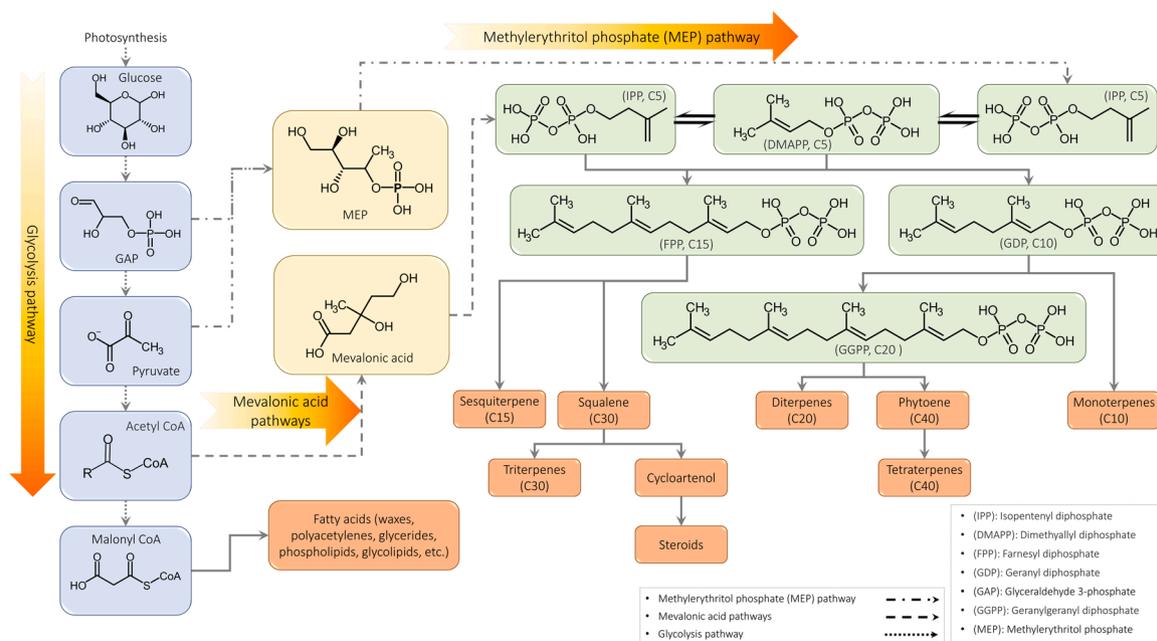
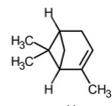
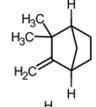
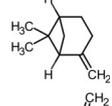
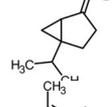
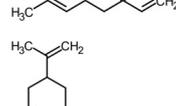
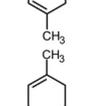
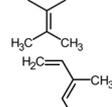
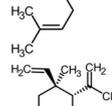
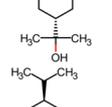
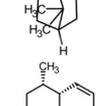
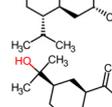
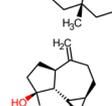
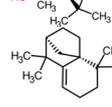
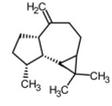
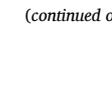


Fig. 2. Biosynthesis of terpene and fatty acid secondary compounds constituting plant resins, showing the interconnections with primary compounds and processes. Elaborated from Wink (2010); Buchanan et al. (2015); Yarnell (2007), and Springob and Kutchan (2009).

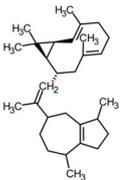
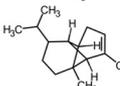
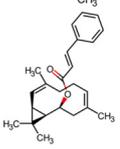
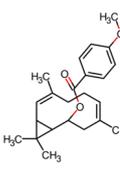
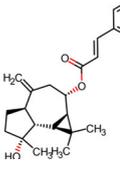
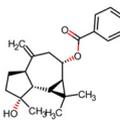
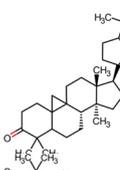
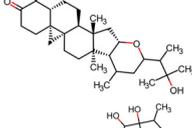
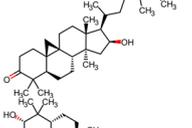
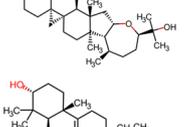
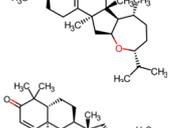
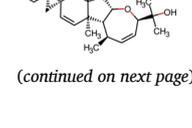
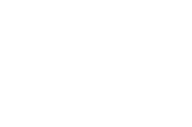
**Table 1**

Guayule resin composition. Compiled from Haagen-Smit and Siu (1944); Scora and Kumamoto (1979); Schloman et al. (1983); Battistel et al. (2017); Parodi (1988); Maatooq and Hoffmann (2002); Saldívar et al. (2003); de Vivar et al. (1990); Gaytán et al. (1997); Banigan and Meeks (1953); Belmares et al. (1980); Punvichai et al. (2016); Mears (1980); Shen et al. (1976); Mabry et al. (1978), and Dehghanizadeh et al. (2020).

Group of compounds	Name	Chemical formula	Composition	Chemical structure
Monoterpenes	$\alpha$ - pinene	$C_{10}H_{16}$	0.5-3%	
	camphene	$C_{10}H_{16}$	<1%	
	$\beta$ - pinene	$C_{10}H_{16}$	0.3-0.7%	
	Sabinene	$C_{10}H_{16}$	0.2-0.5%	
	$\beta$ - myrcene	$C_{10}H_{16}$	~0.1%	
	limonene	$C_{10}H_{16}$	0.2-0.7%	
	terpinolene	$C_{10}H_{16}$	0.3-0.7%	
	$\beta$ - ocimene	$C_{10}H_{16}$	~0.1%	
	Elemol	$C_{15}H_{26}O$	~0.2%	
	Monoterpenoids	l-phellandral	$C_{10}H_{16}O$	~0.2%
borneol <sup>a</sup>		$C_{10}H_{18}O$	<0.1%	
cadinene		$C_{15}H_{26}$	0.3-0.4%	
$\beta$ -eudesmol		$C_{15}H_{26}O$	0.1-0.2%	
Sesquiterpenes and sesquiterpenoids	spathulenol	$C_{15}H_{24}O$	–	
	isolongifolene	$C_{15}H_{24}$	–	
	allo-aromadendrene	$C_{15}H_{24}$	–	
	partheniol	$C_{15}H_{24}O$	–	

(continued on next page)

Table 1 (continued)

Group of compounds	Name	Chemical formula	Composition	Chemical structure
	$\alpha$ -guajene	$C_{15}H_{24}$	~0.1%	
	$\alpha$ -copaene	$C_{15}H_{24}$	—	
	$\alpha$ -humulene	$C_{15}H_{24}$	—	
Sesquiterpene esters	guayulin A	$C_{24}H_{30}O_2$	8-14%	
	guayulin B	$C_{23}H_{30}O_3$	0.5-3%	
	guayulin C	$C_{24}H_{30}O_3$	0.1-3%	
	guayulin D	$C_{23}H_{30}O_4$	0.1-1%	
Triterpenoids	argentatin A	$C_{30}H_{48}O_4$	4-8%	
	argentatin B	$C_{31}H_{50}O_3$	2-4%	
	argentatin C	$C_{31}H_{52}O_4$	~1%	
	argentatin D	$C_{30}H_{50}O_3$	~3%	
	argentatin E	$C_{30}H_{50}O_2$	—	
	argentatin F	$C_{30}H_{42}O_3$	—	

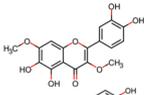
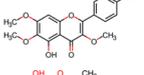
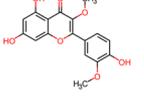
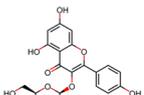
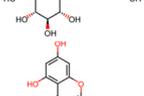
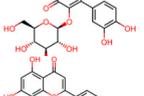
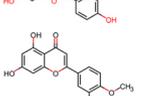
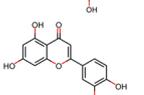
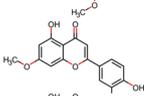
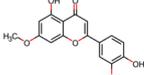
(continued on next page)

Table 1 (continued)

Group of compounds	Name	Chemical formula	Composition	Chemical structure	
	argentatin G	$C_{30}H_{46}O_3$	–		
	argentatin H	$C_{30}H_{48}O_3$	–		
	incanilin (Iso argentatin A)	$C_{30}H_{48}O_4$	–		
	isoargentatin B	$C_{30}H_{48}O_3$	–		
	isoargentatin D	$C_{30}H_{50}O_3$	–		
	argentatin G diacetate	$C_{34}H_{50}O_5$	–		
	argentatin H diacetate	$C_{34}H_{52}O_5$	–		
	Sterols	$\beta$ -sitosterol	$C_{29}H_{50}O$	–	
		trans-cinnamic acid	$C_9H_8O_2$	~4%	
		p-anisic acid	$C_8H_8O_3$	0.5-1%	
palmitic acid		$C_{16}H_{32}O_2$	1.5-2.5%		
stearic acid		$C_{18}H_{36}O_2$	0.3-1%		
Organic acids	oleic acid	$C_{18}H_{34}O_2$	1-2.5%		
	linoleic acid	$C_{18}H_{32}O_2$	10-15%		
	linolenic	$C_{18}H_{30}O_2$	2-4%		
	eicosanoic acid	$C_{20}H_{40}O_2$	<3%		
	heneicosanoic acid	$C_{21}H_{42}O_2$	<1.5%		
	pentadecanoic acid	$C_{15}H_{30}O_2$	<4%		
	Myristic acid	$C_{14}H_{28}O_2$	<2.5%		

(continued on next page)

Table 1 (continued)

Group of compounds	Name	Chemical formula	Composition	Chemical structure
Flavonoids	6-hydroxykaempferol 3,7-dimethyl ether	C <sub>17</sub> H <sub>14</sub> O <sub>7</sub>	–	
	quercetagenin 3,7-dimethyl ether	C <sub>17</sub> H <sub>14</sub> O <sub>8</sub>	–	
	6-hydroxykaempferol 3,6,7-trimethyl ether	C <sub>18</sub> H <sub>16</sub> O <sub>7</sub>	–	
	quercetin 3,3'-dimethyl ether	C <sub>17</sub> H <sub>14</sub> O <sub>7</sub>	–	
	kaempferol 3-glucoside	C <sub>21</sub> H <sub>20</sub> O <sub>11</sub>	–	
	quercetin 3-glucoside	C <sub>21</sub> H <sub>20</sub> O <sub>12</sub>	–	
	Apigenin	C <sub>15</sub> H <sub>10</sub> O <sub>5</sub>	–	
	Diosmetin	C <sub>16</sub> H <sub>12</sub> O <sub>6</sub>	–	
	chrysoeriol	C <sub>16</sub> H <sub>12</sub> O <sub>6</sub>	–	
	luteolin 7-methyl ether	C <sub>16</sub> H <sub>12</sub> O <sub>6</sub>	–	

<sup>a</sup> Derived from hydrolysis of bornyl acetate.

## 2.8. Other compounds

Guayule leaves have been of recent interest as a source of phenolic-rich compounds (Piana et al., 2018; Piluzza et al., 2020). A variety of phenolic compounds and phenolic derivatives, hydroxybenzoic acids, hydroxycinnamic acids, flavanols, flavones, and anthraquinones, have been identified in the leaf biomass (Piana et al., 2018). Five phenolic compounds: neochlorogenic (5-caffeoylquinic acid), chlorogenic (3-caffeoylquinic acid), cryptochlorogenic (4-caffeoylquinic acid), isovanillic, and 3,5-DCQ (3,5-Di-O-caffeoylquinic acid) acids were recently identified in methanol extracts from guayule leaves and twigs (Piluzza et al., 2020). Mears (1980) investigated the distribution of flavonoids in the North American species of *Parthenium* and reported five major flavonoids in guayule (See Table 1).

## 3. Guayule resin extraction

The separation, fractionation, and purification of guayule resin are critical to any value-added use of this multicomponent mixture. Although there have been many proposed commercial applications for guayule resin fractions, few have been economically feasible due to difficulties in the separation and purification of target compounds out of the wide range of molecular weights and polarities. A few studies on separation methods have been conducted for industrial-scale applications. Fractionation of guayule resin has mostly been investigated using conventional solvent extraction techniques. Vacuum and steam distillation have been used for the separation of essential oils, with the

associated thermal degradation of some non-volatile compounds (Haagen-Smit and Siu, 1944; Walter, 1944).

In latex rubber extraction, plant materials are chopped and homogenized in an alkaline aqueous medium using pebble or hammermills, followed by filtering, pressing, washing, and centrifuging to separate latex droplets; there is little extraction of resin. As result, a considerable amount of resin (~8 %) is left in the bagasse or entrained in the rubber phase (Boateng et al., 2010), lowering the initial quality of the rubber and facilitating rubber degradation/oxidation (Jones, 1948).

For dry bulk rubber extraction, resin can be obtained from three methods: flotation, sequential, and simultaneous extraction (Schloman, 2005). In flotation rubber extraction, ground guayule shrub is mixed with a caustic solution to coagulate rubber particles, then the rubber particles are deresinated using a polar solvent (acetone). This method is not suitable for the industrial scale due to the high process water requirements, some energy-intensive steps, the large volume of aqueous waste, and the low quality of produced rubber (van Beilen and Poirier, 2007; Wagner and Parma, 1989). In the sequential extraction method, resin and rubber are extracted from the ground shrub together using a polar solvent (alcohol, ketone, or ester), then rubber is isolated with a non-polar organic solvent. The two-step solvent extraction and the large amount of solvent increase operating costs and potential for solvent loss, making sequential extraction less economically feasible. In simultaneous extraction, rubber and resin are extracted using a mixture of non-polar and polar solvent, for example 20 wt.% acetone and 80 wt.% pentane, to produce a miscella (Cornish et al., 2006; Schloman, 2005); additional polar solvent is added to the miscella to precipitate rubber. Resin is

**Table 2**  
Potential applications of guayule resin.

Category	Application	Description	References
Paints, coatings, and adhesives	Controlled-release formulation	Component of formulations that improve loading and release time of volatile compounds (linalool or carvone), and reduce the porosity of membranes	Pascual-Villalobos and López, 2013
	Resin multipolymers	Reaction of resin with unsaturated monomer for directly use in solvent-less inks, coating, and adhesives	Gumbs, 2009
	Wet-stick bio-adhesives	Bio-adhesives for bonding wet skin, such as pressure-sensitive adhesive tapes, medical tapes, wound or surgical dressings, athletic tapes, surgical drapes, or tapes/tabs for medical sensors	Gumbs, 2008
	Adhesives	Adhesives to bind different kinds of surfaces, even underwater, and to provide insect control	Nakayama, 2005
	Strippable coatings	Combination of guayule resin and epoxy polymers to make strippable coatings for the protection of equipment in storage	Nakayama, 2005
	Composite components	Combinations of wood, plastic, and guayule bagasse or resin for termite- and fungus-resistant products	Nakayama et al., 2004
	Solvent-less adhesives	Combinations of guayule resin with monomers, polymers, and initiators for solvent-less adhesives	Gumbs, 2004
	Material durability enhancers	Ingredient to improve the durability of wood/plastic composites	Chow et al., 2002
	Coatings	Impregnation of woody materials with guayule resin to provide resistance to termite attack and wood-rot	Bultman et al., 1991; Nakayama et al., 2003, 2001
	Coatings	Coatings on wood in marine and terrestrial environments to protect against crustacean borers and termites	Bultman et al., 1998
	Fire retardant	Pine wood impregnation formulation to retard ignition	Smith et al., 1996
	Anti-fouling paints	Anti-fouling paints for marine applications that are less toxic to shrimp and/or barnacles	Thames et al., 1996
	Plasticizer/adhesive	Plasticizer/adhesive modifiers in high-performance epoxy resin coatings	Thames and Kaleem, 1991 Thames and Kaleem, 1991; Thames and Wagner, 1991
	Strippable coatings	Adhesive modifier for strippable coatings	
	Antifungal, anticancer, and antimicrobial agents	Plasticizer for high polymers	Plasticizer for tailoring stress-strain properties
Coatings		Water and abrasion-resistant coatings	Belmares et al., 1980
Antiplastic agent		Guayulin A and B as biological triggers for synthesizing antiplastic agent in breast cancer treatment	(Spano et al., 2018)
Anti-cancer agent		Argentatin A and B are cytotoxic and cytostatic, respectively, for proliferating lymphocytes.	Parra-Delgado et al., 2005
Anti-cancer		Argentatin B inhibits the proliferation of human colon cancer cell line (RKO)	Romero-Benavides et al., 2018
Antitumor agent		Anticancer activity of argentatin A in HCT116 colon cancer cells	Tavarez-Santamaría et al., 2020
Antitumor agent		Argentatin B inhibits the proliferation of cancer cells (HCT-5 and PC-3)	Alcántara-Flores et al., 2015
Antifungal		Sesquiterpenoid extracts used to treat fungal infections	Maatooq and Hoffmann, 1996; Maatooq et al., 1996
Antimicrobial		Argentatin A activity against various microorganisms	Martínez et al., 1986; Martínez Vazquez et al., 1994
Antineoplastic agent		Guayulin A and B for synthesizing antineoplastic agent lychnostatin 1	Zoeller et al., 1994
Antitumor agent taxol		Guayulin A and B for anticancer treatments	Zoeller et al., 1994
Anticancer agents		Argentatin A, B, and its derivatives for cytotoxic and anti-inflammatory treatments	Parra-Delgado et al., 2005
Termite repellent		Natural and safe termiticide	Bajwa et al., 2017
Bio-control agents		Argentatin A and B for growth inhibition of the fall armyworm <i>Spodoptera frugiperda</i>	Carlos et al., 2001
Pesticides		Termite control agent	Sesquiterpenes and triterpenes as insect anti-feedants and toxins
	Cockroach pheromone	Guayulin A and B for synthesizing cockroach pheromone periplanone B	Zoeller et al., 1994
	Bio-control agents	Guayule resin negative impacts on larva stage of certain insects	Isman and Rodriguez, 1983
	Softener	Guayule resin as a softener in the tire sealant composition	Randall et al., 2020
	Epoxidized natural rubber	Epoxidized natural rubber from LMWR fractions to enhance physical properties of cured rubber	Schloman, 1992
Rubber additives	Rubber additives	Sulfurized resin to reduce hysteresis loss and increase tensile strength in conventional rubber	Schloman and Davis, 1986
	Tackifying agents	Green tackifiers and strength enhancers for improving rubber	Kay and Gutierrez, 1984, 1985
	Peptizing agent	Peptizing agent for both <i>Hevea</i> natural rubber and styrene/butadiene rubber	D.S. Winkler, 1977
Fuels	Liquid fuels	Conversion of guayule resin to liquid fuel by cracking catalysis	Gutierrez and Kay, 1983
	Biofuel feedstock	Mixture of guayule bagasse and resin	Sabaini et al., 2018
	Liquid hydrocarbons	Conversion of guayule resin to C <sub>1</sub> -C <sub>10</sub> hydrocarbons using zeolite-based catalysts	Costa et al., 1992
	Asphalt binder modifier	Guayule resin as recycling agent in asphalt binders	Hemida and Abdelrahman, 2019; Lusher and Richardson, 2015
Miscellaneous	Soil amendment	Resin-containing bagasse as soil amendment to improve lettuce growth	Schloman, 1991
	Essential oils	Essential oil from steam distillation of fresh guayule biomass for fragrance/volatiles applications	Haagen-Smit and Siu, 1944

recovered from the miscella by evaporating the solvent. The main advantage of this method is that the quality of the final rubber product, particularly the rubber viscosity and resin content, can be adjusted (Beinor and Cole, 1986). This capability is important for managing variations between guayule shrub lines and harvested biomass

properties. The main disadvantages of this method are the complex separation processes for recycling solvents and removing the solvent from the rubber after precipitation (Wagner and Parma, 1988). Resin yield from sequential extraction is significantly higher (68 %) than from simultaneous extraction, however, the resin is noticeably heterogeneous

and contains entrained water-soluble materials, including 1.4 wt.% ash; resin from simultaneous extraction is homogeneous and contains no water or water-soluble materials (Schloman et al., 1988). The presence of water and water-soluble materials complicate the solvent recovery process, and lower rubber yield and quality, therefore, guayule biomass is sometimes rinsed with water then dried before extraction (Black et al., 1986; Curtis, 1947). Although total guayulin, argentatin, and triglyceride contents in resin from sequential extraction are lower than those in resin from simultaneous extraction, the concentration of reactive groups is higher, increasing the potential for chemical modification with polyamines or amine-terminated polyethers (Schloman, 1988a, 1988b).

Supercritical CO<sub>2</sub> extraction has been investigated as an alternative for the guayule resin and rubber separation (Cornish et al., 2006; Punvichai et al., 2016). The extraction was significantly faster and better controlled than conventional liquid solvent extraction. Parameters that significantly affected the extraction yield and selectivity were pressure (10–69 MPa), temperature (60–100 °C), co-solvent ratio, and time (Cornish et al., 2006). Some (2–3 wt.%) resin remained in shrub biomass after supercritical extraction with pure CO<sub>2</sub> (Cornish and Marentis, 2010). Small amounts of co-solvent can significantly promote resin and rubber extraction yield; biomass particle size, moisture content, and pretreatments also play important roles in extraction efficiency. The highest yield of resin was obtained at 45–60 °C, 200–300 bar, and 60 min, using ethanol as the co-solvent (Cornish et al., 2006). Supercritical CO<sub>2</sub> extraction has been used for pretreating guayule bagasse to remove resin prior to its use as a feedstock for enzymatic hydrolysis (Srinivasan and Ju, 2010). A summary of these supercritical CO<sub>2</sub> extraction experiments is shown in Table A1.

Accelerated solvent extraction (ASE) has been investigated for guayule rubber and resin extraction at the lab scale (Cornish et al., 2013; Pearson et al., 2013; Suchat et al., 2013). Resin was extracted with a polar solvent at lower temperatures and the rubber was recovered using a non-polar solvent at higher temperatures. The maximum yield of resin and rubber (from a two-year-old shrub, oven-dried at 50 °C) was obtained after three rinses with acetone at 40 °C and three rinses with hexane at 140 °C, respectively (Pearson et al., 2013). Drying temperatures above 75 °C can degrade rubber into the acetone phase during ASE, thus overestimating resin and underestimating rubber contents (Black et al., 1983; Cornish et al., 2013; Pearson et al., 2013). Experimental results for ASE of guayule resin and rubber are summarized in Table A2.

Some other alternative techniques, including electrostatic separation, adsorptive separation, microfiltration, ultracentrifugation, and liquid cyclones have been proposed for resin extraction and purification. Clogging and high maintenance costs are the main barriers to commercialization (Thames and Wagner, 1991). The feasibility of using ultrafiltration (UF) membranes for single-step fractionation of guayule resin has been demonstrated by Jeyaseelan and Wagner (1995) and Wagner and Parma (1989). The solvents (xylene and methanol) and lower-molecular-weight fatty acids (C<sub>16</sub>–C<sub>18</sub>) are separable in a single-step ultrafiltration system using a ceramic membrane (Wagner and Parma, 1989). The ambient operating temperatures in ultrafiltration prevent the degradation of thermolabile constituents of guayule resin. Volatile essential oils can be separated from resin using cellulose triacetate membranes with 200 and 500 Da molecular weight cut offs, however, low flow rates and membrane fragility limit the commercial application of the technique (Daly, 1989; Jeyaseelan and Wagner, 1995).

Efficient separation and fractionation of guayule resin (or other resin-containing plants) with high chemical complexity and significant differences in compound polarity may require a combination of separation techniques. For example, using ultrafiltration alone may not be economically feasible for large-scale applications, but in combination with supercritical fluid extraction (SFE) or pressurized liquid extraction (PLE), the production cost can be significantly decreased (Sarmiento et al., 2008, 2004; Spricigo et al., 2001). Selection of individual and/or combination techniques will all need to consider the associated economic tradeoffs, many of which have not yet been studied.

#### 4. Guayule resin applications

Value-added use of guayule processing residues has the potential to reduce gross rubber production costs by 26–49 % (Schloman et al., 1986). Wright et al. (1991) suggested that research and development into such applications might even create the scenario where co-products derived from guayule resin, LMWR, and/or bagasse exceed the value of the rubber itself (Nakayama, 2005). Potential direct uses of crude guayule resin (that do not require pharmaceutical isolation and purification) include coatings, adhesives, viscosity modifiers, and plasticizers (Schloman and Wagner, 1991). Recent focus on economics has led to developments in guayule resin applications in coatings, biocontrol agents, and controlled-released formulations (Pascual-Villalobos and López, 2013). Argentatins have been reported to have antimicrobial (Martinez Vazquez et al., 1994), insecticidal (Carlos et al., 2001), and antitumoral properties (Alcántara-Flores et al., 2015). Polyphenolics, cinnamyl derivatives, and terpenoids are the most likely compounds responsible for the observed termite resistance characteristics of guayule resin (Bultman et al., 1986). The volatile fractions of guayule resin are rich in alpha-pinene, beta-pinene, and limonene—of which 30,000 tons are produced annually for fragrances and adhesives (Swift, 2004). The recently-identified phenolic compounds in the leaves and twigs have potential uses in antioxidant, anticancer, anti-inflammatory, and anti-diabetic applications (Piluzza et al., 2020). Table 2 provides a summary of guayule resin applications that have been reported but not yet commercialized.

#### 5. Guayule resin economics

The economic feasibility of guayule rubber extraction is highly sensitive to the raw material value of the co-products (Sproul et al., 2020). The bagasse represents a relatively fixed revenue potential as it is dried biomass without an exotic compound. The resin represents a unique opportunity based on the diversity of compounds contained. The first step to estimating the resin's raw material value is to understand the range of active ingredients/products that resin can replace. Among the product replacements studied have been bio-control agents and controlled released formulations (Pascual-Villalobos and López, 2013); coatings, adhesives, viscosity modifiers, and plasticizers (Schloman and Wagner, 1991); insecticides (Gutiérrez et al., 1999); and wood preservatives (Bultman et al., 1991; Nakayama et al., 2003, 2001) (Table 2). To down-select from the long list of potential guayule resin uses, retail prices (USD/kg) and market size of key product categories were estimated using published literature and online marketplaces. Bulk retail prices, technical cost reports, and financial marketplace websites were prioritized as the most appropriate sources for wholesale product prices. When information from these sources was not available, online retail prices were used for approximations. Product categories included asphalt binders, adhesives, rubber tackifying resins, paints and coatings, soil amendments, and essential oils (Tables A3 and A4). Using this data, a landscape of potential guayule resin product prices was created (Fig. 3).

The distribution in Fig. 3 shows the resin can be used in high- and low-price products. In general, the low-price products have the largest market sizes. Conversely, products with the highest retail price are typically for smaller market sizes: antifouling paints, essential oils, marine wood coatings, wood preservative coatings, and pesticides. “Paints and Coatings” was added to account for the wider variety of coating types and paints with resin applications. Asphalt binders, soil amendments, and rubber tackifying resin are lower in price, while solvent-based, solventless, and medical spray adhesives have intermediate prices. The wide range of specific prices is attributed to variation based on brand, size, type, and specific function. Additionally, there is uncertainty about the fraction of the product that can be replaced by the resin and the extent to which the resin needs to be fractionated (at additional cost of manufacturing).

A combination of high price and moderate market size suggests the

products with the highest economic potential (without accounting for processing costs for the raw material) are residential pesticides, paints and coatings, wood preservative coatings, amine-epoxy strippable coatings, and adhesives. These five most promising resin co-products were investigated further. Fig. 4 shows resin prices estimated according to the resin's function and mass fraction in each product. A distinction between conservative, baseline, and optimistic prices was made based on the range of prices found while surveying each product. The baseline estimates are based upon a single source value that represents a central estimate within the range. The conservative and optimistic prices are the minimum and maximum values of the prices found. Including conservative, baseline, and optimistic prices for each category conveys the uncertainty associated with these initial estimates.

Within Fig. 4, mass fractions of the resin were applied to the overall product prices to estimate the portion of the price that can be attributed to the resin. For pesticides, wood preservative coatings, and paints and coatings, resin accounts for 10–40 % of the total product composition (Table A5), assuming the insecticidal/anti-termitic function of the resin can entirely replace the typical fraction of active ingredients. For adhesives, resin accounts for 5–40 % of the total product assuming resin acts as the primary base resin in polyurethane, hot melt packaging, water born polyurethane, and pressure sensitive adhesive formulations (Strickland, 2013). For amine-epoxy strippable coatings, resin can account for up to 10 % of the total product as a plasticizer and maintain performance characteristics (Thames and Kaleem, 1991). Based on these application assumptions, guayule resin has an estimated price ranging from 0.1 to 6.8 USD/kg. Domestic pesticide use gave the highest specific price (1.1–6.8 USD/kg), followed by wood preservative coatings (0.8–6.0 USD/kg), paints and coatings (0.7–6.0 USD/kg), and epoxy strippable coatings (0.8–2.5 USD/kg). The specific price was lowest for adhesive applications (0.125–1.6 USD/kg). It is important to note that these price ranges do not account for the fractionization, processing, packaging, and distribution of the resin and/or final products. Further modeling and experimental work are required to fully understand the raw material value for resin.

## 6. Recommendations for guayule resin biorefining

The complexity of natural resin means that there can be no single technique for separation. Previous studies have concentrated on identifying the most efficient extraction method for specific groups of natural resin compounds based on chemistry. The next step for guayule resin

commercialization is identifying a series of efficient separation techniques in a reliable chemical process model that can be used to inform economic and life cycle assessments. A potential guayule resin refinery model, with alternative pathways, is proposed in Fig. 5 based on previous characterization work (Cheng et al., 2020; Dehghanizadeh et al., 2020). This model was based on two scenarios: (1) freshly-harvested and (2) field-dried shrub. Output stream decision points are shown as diamonds, allowing multiple scenarios to be shown on a single diagram. Unit operation selection is based on three factors: a) the potential for scale up, b) the extraction efficiently for target compounds, and c) minimization of environmental impacts. Pretreatment steps, like parboiling, milling, grinding, de-leafing, stabilizing, hydrolyzing, and flaking, were not included in the model and would need to be considered in a final evaluation. Such pretreatment steps can and do strongly affect the efficiency of processing steps.

From Fig. 5, the first unit (110) for fresh shrub material is an essential oil (EO) extraction such as hydro-distillation or steam distillation. Non-volatile compounds from EO extraction can either be sent to a drying unit (130) or to a microwave-assisted extraction (MAE) separation unit (120), which can handle wet biomass after adjusting the moisture content of the solid-liquid mixture with a dewatering unit. Microwave-assisted extraction would remove the majority of guayule resin constituents (sesquiterpenes, triterpenes, phenolics, and fatty acids). Residues, rich in rubber and bagasse, would then be sent to the drying unit (130). This pathway is expected to slightly increase the rubber concentration in the non-volatiles residues and act as a pretreatment step, since the walls of the rubber-containing cells would be disrupted by microwaves. Due to the poor selectivity (presence of LMWR in acetone extracts and resin in hexane extracts) and environmental concerns associated with using organic solvents in conventional solvent extraction, SFE/PLE (units 140 and 150) would be used next as highly selective methods to separate the wide range of secondary metabolites, with rubber and bagasse being products after unit 140. A similar process using supercritical CO<sub>2</sub> extraction in separation, fractionation, and purification of rubber and resin was patented by Cornish et al. (2006).

The first scenario was developed to exploit the maximum amount and quality of terpenoids (e.g.  $\alpha$ -pinene,  $\beta$ -pinene, caryophyllene, camphene, etc.). The yield and composition of major volatile compounds are strongly influenced by the drying process (Dong et al., 2011; Ghasemi Pirbalouti et al., 2013; Szumny et al., 2010). Similarly, some hydrocarbons are volatilized due to the heat produced during milling. Field-dried biomass has the advantage of being easily transported,

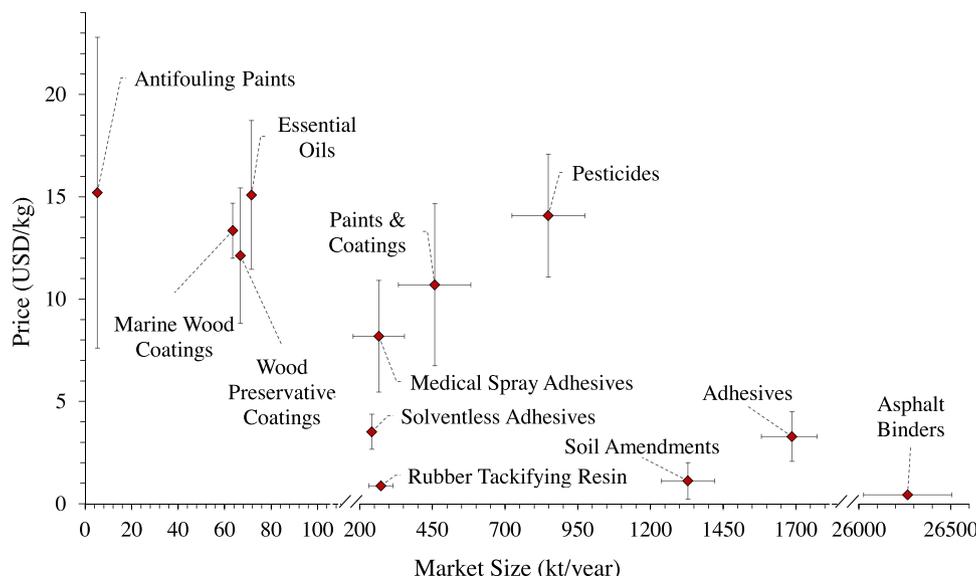


Fig. 3. Estimated prices of potential resin-containing products; uncertainty bars represent the range of prices reported in the references surveyed.

stored, and handled; on the other hand, freshly-harvested shrub has the advantages of avoiding the biochemical reactions and increased biological deterioration of aromatic plants that occurs during drying (Díaz-Maroto et al., 2003; Hamrouni-Sellami et al., 2013). The total amount of guayulins A–D in a fresh sample is approximately 15 % higher than in dried samples, a factor that cannot be ignored if applications of the resin sesquiterpenes are to be targeted (Spano et al., 2018). Assuming guayulins C and D are most likely derived from the oxidation/degradation of guayulins A and B, the concentration of guayulins C and D were significantly higher in the dried shrub (Martínez et al., 1986; Spano et al., 2018). Guayule leaves (green and dried) and flowers (peduncles and inflorescences), which contribute on average 32 % and 6 %, respectively, to the total biomass weight (Teetor et al., 2009) and are rich in resin (7–8 %), protein (12–22 %), and waxes (2.5 wt.% dry basis) (Banigan et al., 1982), are discarded prior to bulk rubber extraction. Wax extracted from guayule leaves has one of the highest melting points (76 °C) recorded for a natural wax, giving it value-added potential as a source of hard wax (Banigan et al., 1951). Since guayule rubber is also synthesized and stored in the mesophyll cells of leaves (Madhavan and Benedict, 1984), the presence of leaves, which contain 0.5–2.0 % of total plant rubber (Gilliland and van Staden, 1986; Teetor et al., 2009), can increase the total rubber yield. Therefore, processing of fresh guayule shrub provides the opportunity to extract more terpenes, protein, and waxes than field-dried shrub.

Latex extraction (unit 300) from fresh shrub is another approach to rubber extraction since the bagasse produced from latex extraction is rich in resin (around 40 %) that may be a suitable feedstock for the SFE. Traditional resin extraction from resinous bagasse is difficult and inefficient because of the low percolation rate of solvents (Cornish et al., 2006). Supercritical CO<sub>2</sub> extraction (in unit 150) with high diffusivity properties has the potential to penetrate the low porosity bagasse cells and facilitate the release of compounds of interest at high percolation rates. Supercritical CO<sub>2</sub> with ethanol co-solvent can increase resin yield up to 12 % of total defoliated biomass compared with 7 % by acetone extraction (Punvichai et al., 2016).

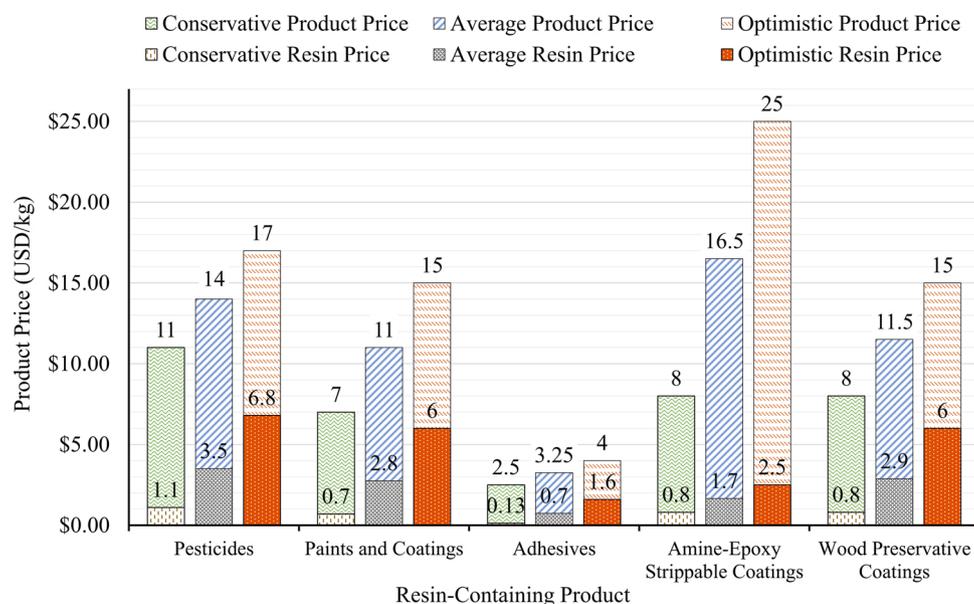
The scenario using field-dried shrub, starting with unit 200, addresses the difficulties of using fresh shrub: fungal contamination, mold and bacteria growth, limitations caused by the presence of moisture on some extraction methods like SFE, and the presence of leaf material. (Leaf materials can reduce the rubber recovery due to the formation of

very fine particles that clog the filtration systems and increase the ash content associated with rubber degradation and quality reduction (Battistel et al., 2017).) The bagasse produced from simultaneous extraction (unit 210) can be further processed with supercritical CO<sub>2</sub> to remove the residual solvent and extract value-added chemicals (pathway not shown). Key to the proposed dried shrub pathway is the multiple options for resin separation immediately after rubber processing, including SFE/PLE (unit 140), solvent extraction (unit 230) followed by SFE/PLE or terpene fractionation (unit 240), or saponification (unit 250) followed by solvent extraction or vacuum distillation (unit 260). Each pathway leads to similar groupings of resin products and could be tuned depending on the end product markets.

## 7. Conclusions

This review provides insight into the chemistry, extraction methods, and applications of guayule resin. The complexity and range of polarity of molecules in guayule resin mixtures (terpenoids, fatty acids, phenolics, steroids, alkaloid compounds, etc.) necessitate the evaluation of many methods for extracting constituents for value-added applications. Characterizing the distribution of chemical structures in guayule resin is a crucial step for finding the most appropriate extraction method. Modern extraction techniques reduce extraction time, energy, and solvent consumption, yet a completely “green” technique that enables isolation of all of the targeted compounds simultaneously is still elusive.

Guayule resin, often considered in the context of a byproduct of guayule rubber production, has the potential to overtake rubber value if the resin fractions can be suitably separated and used in high-value, medium-scale applications such as pesticides, paint, and coatings. Of the developed methods for natural resin fractionation with potential for scale-up, pressurized solvent extractions, like supercritical CO<sub>2</sub> and pressurized liquid extraction, are advantageous over other extraction methods due to the increased solubilizing power for the desired compounds and the higher solvent diffusivity. Care must be taken with higher extraction temperatures to avoid degradation of some high-value thermolabile compounds and biopolymers (rubber) into the resin extract phase. The potential of resin utilization for improving the economic viability of guayule as an industrial crop means that economic and lifecycle analyses of conceptual biorefineries need to include multiple pathways, starting from the whole plant.



**Fig. 4.** Potential products and estimated resin prices based on the respective resin functions. Price bars are divided into product (lighter color) and the mass fraction that is assumed to be replaced with resin (darker color).

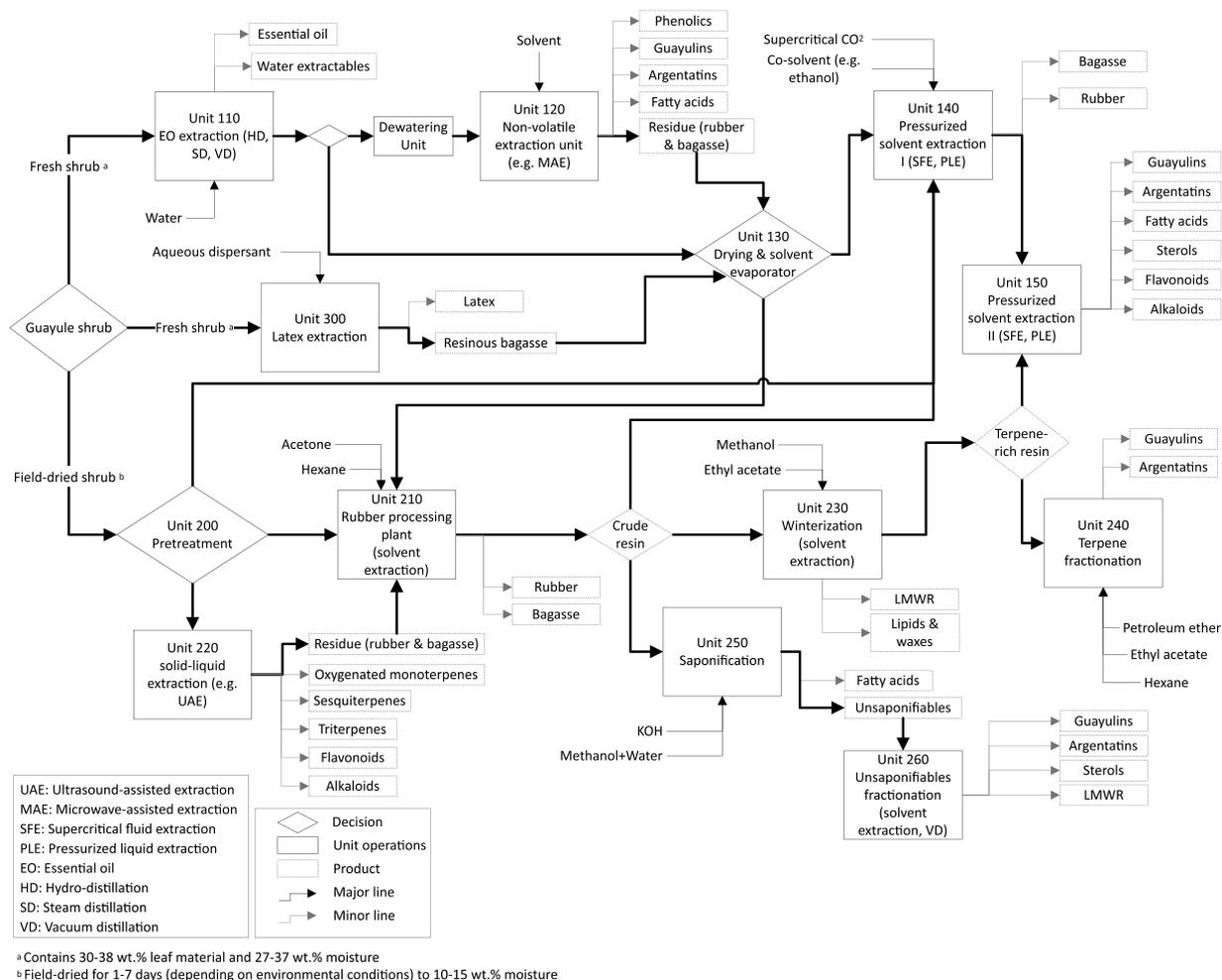


Fig. 5. A block flow diagram (BFD) of proposed guayule resin refinery processes.

**Declaration of Competing Interest**

The authors report no declarations of interest.

**Acknowledgments**

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**Appendix A**

**Table A1**

Guayule resin and rubber extractions using supercritical CO<sub>2</sub> extraction, summarized from Cornish et al. (2006) and Punvichai et al. (2016).

Sample	T (°C)	P (bar)	time (min)	Co-solvent (co-solvent/feedstock ratio)	CO <sub>2</sub> flowrate	Extract yield (wt.%)	Results
1 <sup>a</sup>	60	345	30	–	3 (liters/min)	2.9	High selectivity for resin
2 <sup>a</sup>	100	675	23	hexane (4/1)	3 (liters/min)	8.0	Best extraction capability for rubber and lowest concentration of resin
3 <sup>a</sup>	100	345	45	hexane (22/3)	3 (liters/min)	4.7	Good extraction capability for rubber and low Resin concentration
4 <sup>a</sup>	80	675	60	hexane (17/2)	3 (liters/min)	11.2	Very good selectivity for rubber
5 <sup>a</sup>	100	675	60	1-hexane (17/2)	3 (liters/min)	5.9	Very good selectivity for rubber
6 <sup>a</sup>	80	655	60	cyclohexane (17/2)	3 (liters/min)	6.2	Very good selectivity for resin
7 <sup>a</sup>	100	655	60	iso-octane (17/2)	3 (liters/min)	5.5	Good selectivity for resin
8 <sup>a</sup>	100	675	30	water (1/2)	3 (liters/min)	4	High selectivity for resins and relatively low selectivity for rubber
9 <sup>a</sup>	60	345	24	water (1/2)	3 (liters/min)	4.2	Concentration of resin is not nearly as high as the previous experiment. The yield is highest
10 <sup>a</sup>	9.2	138	–	–	–	–	No extract is collected

(continued on next page)

Table A1 (continued)

Sample	T (°C)	P (bar)	time (min)	Co-solvent (co-solvent/ feedstock ratio)	CO <sub>2</sub> flowrate	Extract yield (wt.%)	Results
11 <sup>a</sup>	60	345	45	acetone (1/1)	3 (liters/min) 3 (liters/min)	4.9	Extraordinary selectivity for resin.
12 <sup>b</sup>	40	250	60	acetone	34.4 (g/min)	4.8	After 60 min increasing extraction time doesn't results in a visible gain. (Yield <sub>120 min</sub> = 5.1 %)
13 <sup>b</sup>	33	250	60	acetone	34.4 (g/min)	6.3	This yield is higher than the obtained yield (5.2 %) from ASE (ref. Method) using acetone
14 <sup>b</sup>	33	250	60	ethanol	34.4 (g/min)	9.6	Ethanol clearly displays a better co-solvent power than acetone
15 <sup>c</sup>	33	250	60	ethanol	34.4 (g/min)	10.1	Optimum flowrate of CO <sub>2</sub> : 34-51 ml/min
16 <sup>c</sup>	33	200	60	acetone	34.4 (g/min)	7.7	No significant changes in yield with increasing "P" from 200 to 300 bar using acetone as co-solvent
17 <sup>c</sup>	33	300	60	ethanol	34.4 (g/min)	12.1	A constant increase in yield with increasing "P" from 100 to 300 bar using ethanol as co-solvent
18 <sup>c</sup>	60	250	60	ethanol	51.6 (g/min)	12.3	Maximum yield is obtained between 45–60 °C using ethanol as co-solvent
19 <sup>c</sup>	60	250	60	acetone	51.6 (g/min)	7.03	Co-solvent power of acetone is almost T independent over the tested range (33–80 °C)

<sup>a</sup> Feedstock: Entire fresh sample, chipped: 3/8", dried at 80 °C.

<sup>b</sup> Feedstock: defoliated shrub, vacuum-dried at 40 °C for 3 days. Moisture <10 %. Harvest place: France.

<sup>c</sup> Feedstock: Bagasse obtained after water-based extraction. Oven dried at 40 °C. Ground 0.5 mm. Harvest place: Spain.

Table A2

Guayule resin and rubber extraction using ASE (accelerated solvent extraction).

Sample	Solvent	Cycle/time (min)	T (°C)	Resin yield (wt.%)	Solvent	Cycle-time (min)	T (°C)	Rubber yield (wt.%)	Remark	Reference
1 <sup>a</sup>	acetone	repeatedly/30	100	7.2	–	–	–	–	–	Pearson et al. (2013)
2 <sup>a</sup>	acetone	three/20	100	9.4	cyhex	three/20	140	4.0	–	
3 <sup>a</sup>	acetone	two/5	40	7.4	cyhex	three/20	140	5.3	–	
4 <sup>b</sup>	acetone	three/5	100	8.0	hex	three/20	140	2.3	0 h. pre-extraction heating time at 200 °C	Cornish et al. (2013)
5 <sup>b</sup>	acetone	three/5	100	3.2	hex	three/20	140	0.1	12 h. pre-extraction heating time at 200 °C	
6 <sup>c</sup>	acetone	three/5	100	12.4	hex	three/20	140	0.8	1 h. pre-extraction heating time at 150 °C	
7 <sup>c</sup>	acetone	three/5	100	12.6	hex	three/20	140	0.8	0.3 h. pre-extraction heating time at 200 °C	Suchat et al. (2013)
8 <sup>d</sup>	acetone	three/5	100	9.4	cyhex	three/20	140	7.1	48 h. pre-extraction heating time at 50 °C	
9 <sup>d</sup>	acetone	three/5	100	9.5	cyhex	three/20	140	7.6	24 h. pre-extraction heating time at 75 °C	
10 <sup>e</sup>	acetone	three/5	100	5.5	cyhex	three/20	140	6.9	4 h. pre-extraction heating time at 75 °C	Salvucci et al. (2009)
11 <sup>f</sup>	acetone	three/5	100	16.2	cyhex	two/20	100	1.5	–	
12 <sup>g</sup>	acetone	two/20	24	8.8	cyhex	two/20	100	7.3	air-dried 4 days at 23 °C	
13 <sup>g</sup>	acetone	two/20	24	6.2	cyhex	two/20	100	8.4	air-dried 4 days at 23 °C	Taurines et al. (2019)
14 <sup>h</sup>	acetone	three/20	40	9.3	hex	three/25	120	9.05	–	
15 <sup>i</sup>	acetone	three/20	40	10.7	hex	three/25	120	5.8	–	
16 <sup>j</sup>	acetone	three/20	40	9.3	hex	three/25	120	9.2	–	

<sup>a</sup> 2-year old shrub, AZ2 line, hand-harvested, foliated, dried at 65 °C.

<sup>b</sup> bagasse from latex extraction of 3-4-year-old shrub in aqueous hydroxide, partially defoliated, dried outdoors at AMT, ground for 90 s using 2-mm mesh screen.

<sup>c</sup> 2-year old shrub, AZ2 line, hand-harvested, foliated, dried at 50 °C, ground for 90 s using 2-mm mesh screen.

<sup>d</sup> Three-year old shrub, defoliated, harvested in summer.

<sup>e</sup> Three-year old, collected during autumn.

<sup>f</sup> Guayule stem tissue with high latex rubber content (~9.6 %), harvested from plant grown under 9 h, 16°C light/15 h, 7°C dark photoperiod, freeze-dried at -80 °C.

<sup>g</sup> Two-year old shrub, AZ2 line, defoliated, ground to 1.7 mm, latex content.5.3 %.

<sup>h</sup> 1-2-year-old shrub, harvested in Spain from AZ2, AZ1, 11595, N565, 593, and CAL-6 lines, leaves and flowers, partially dried at AMT for two days, ground to <0.5 mm, vacuum dried at 40 °C for 48 h.

<sup>i</sup> ~ 4-year-old shrub, line CL1 (parent 11591, triploid), leaves and flowers removed, dried at 70 °C for 15 h, freeze dried at -80 °C for 15 h, ground to 1 mm.

<sup>j</sup> ~ 4-year-old shrub, line CLA1 (parent AZ 101, tetraploid), leaves and flowers removed, dried at 70°C for 15 h, freeze dried at -80 °C for 15 h, ground to 1 mm.

**Table A3**  
List of potential products and the corresponding economic value.

Product	Value Range (USD/kg)	Source	Item
Asphalt binder	0.26-0.61	The Ohio Department of Transportation, 2019	
		Poten and Partners Asphalt Weekly Monitor 2017	
		Colorado Asphalt Pavement Association 2012-Present	
		Idaho Department of Transportation Asphalt Price Index 2020	
		Oregon Department of Transportation Monthly Asphalt Cement Material Price 2019	
		Louisiana Department of Transportation and Development 2019	
		New Mexico Department of Transportation 2019	
		Arizona Asphalt Price Index 2019	
		American Floor Mats	Rubber Flooring Adhesive Item No: STADHRFA
		Zoro	Roberts #3000-4 ZORO #: G9189680
Adhesives	1.30- 9.03	ACE Hardware	Zinsser SureGrip 122 High Strength Adhesive 1 gal.
		Zoro	Rust-Oleum #02880 Zoro #: ZORO #: G9489234
		Zoro	Catchmaster #BG-1 ZORO #: G3279245
		Zoro	Weldwood #00142 ZORO #: G2310366
		Lumber Liquidators Flooring Company	Pro Bond Adhesive 5 Gallon SKU: 10047804
		Lumber Liquidators Flooring Company	Ultrabond eco 995 Adhesive 5 Gallons SKU: 10039742
		Zoro	Roberts #1407-1 ZORO #: G5123727
		Zoro	Roberts #2057-4 ZORO #: G9189568
		Zoro	Roberts #2057-1 ZORO #: G5118233
		The Home Depot	Roberts 4 Gal. Multi-Purpose Carpet and Sheet Vinyl Adhesive (24 Pail Pallet) Internet #202935530
Solventless (laminating) adhesives	2.07- 7.88	EMI Supply	Titebond Solvent Free Multi-Purpose Flooring Adhesive, 1 Gallon Can
		The Home Depot	Roberts 4 Gal. Premium Vinyl Tile Glue Adhesive (24 Pail Pallet)
		The Home Depot	Henry 4.75 Gal. 203 Cold Applied Roof Adhesive
		The Home Depot	Roberts 4 Gal. Resilient Flooring Adhesive for Fiberglass Sheet Goods and Luxury Vinyl Tile
		The Home Depot	Custom Building Products SimpleSet Gray 1 Gal. Premixed Thin-Set Mortar
		The Home Depot	Titebond III Ultimate Wood Glue (2-Pack)
		The Home Depot	OmniGrip 3-1/2 Gal. Maximum Strength Adhesive
		The Home Depot	Custom Building Products AcrylPro 3-1/2 Gal. Ceramic Tile Adhesive (24 buckets/pallet)
		The Home Depot	Roberts 1 Gal. Universal Flooring Adhesive
		Reportlinker. "The Laminating Adhesives Market Is Projected to Register a CAGR of 6.9 %, in Terms of Value, between 2019 and 2024." <i>PR Newswire: Press Release Distribution, Targeting, Monitoring and Marketing</i> , 21 Oct. 2019, <a href="http://www.prnewswire.com/news-releases/the-laminating-adhesives-market-is-projected-to-register-a-cagr-of-6-9-in-terms-of-value-between-2019-and-2024-300941835.html">www.prnewswire.com/news-releases/the-laminating-adhesives-market-is-projected-to-register-a-cagr-of-6-9-in-terms-of-value-between-2019-and-2024-300941835.html</a> .	
Medical spray adhesives	5.45- 10.87	"Spray Adhesives Market." <i>Market Research Firm</i> , Markets and Markets, Jan. 2018, <a href="http://www.marketsandmarkets.com/Market-Reports/spray-adhesive-market-201133268.html">www.marketsandmarkets.com/Market-Reports/spray-adhesive-market-201133268.html</a> .	
Rubber tackifying resin	0.75- 2.09	Southern Geo Supply	Profile Tornado Tack ST-1000 Straw Tackifier, 50 lb Bag SKU: ST-1000
		Southern Geo Supply	Profile Tacking Agent 3, Tackifier and Dust Control SKU: LF21TACK3
		Alibaba	Rubber Chemicals Petroleum Resin Tackifier C5 CAS- No: 64742-16-1
		Alibaba	Rubber Chemicals Petroleum Resin Tackifier C5 CAS- No: 64742-16-2
Strippable coatings	9.68- 27.18	C&C Industrial Sales	BINKS Part No: 29-249 Booth Coat (White)
		TOOLiD	3M® 6840 - 5 Gal. Booth Coating Item # mpn1453059886
		Zoro	Precision Brand #43125 ZORO #: G7065307
		MSC Direct	Clear Water Base Booth Coating Part #:00240069
		Collisions Services	Klean-Strip Mask & Peel Spray Booth Coating CMP229 SKU #1700850
		The Home Depot	Olympic Waterguard 1 gal. Clear Multi-Surface Waterproofing Sealant SKU #1003317178
		Amazon	Rust-Oleum 1904A Wolman (Woodlife) CopperCoat Green Wood Preservative-Below Ground, Quart 1904A
		Amazon	Rust-Oleum Clear 902 Wolman Classic Wood Preservative-Above Ground, Quart Part Number 902
		Amazon	Watco 68141 Wipe-On Polyurethane Finish, Quart, Clear Satin Part Number 68141
		ACE Hardware	Flood CWF-UV Matte Redwood Water-Based Wood Finish 5 gal. Item no.1465160
Wood preservative coating & Marine wood coating	2.90- 17.16	Amazon	DEFY Extreme 5 Gallon Semi-Transparent Exterior Wood Stain, Natural Pine Part Number 300163
		East Coast Hardware	Twp 1500 Series Twp-1501-1 Wood Preservative, Cedartone, 1 Gal Can SKU# 5111515
		Amazon	Gemini 204082 TWP1500-1 1 G Clear Wood Preservative Part Number 204082
		Green Building Supply	Vermont Natural Coatings, Exterior, Penetrating Water Proofer Item #42075
		Western Log Home supply	X-100 Natural Seal Wood Protective for Log Homes Item #: NSWP5
		Walmart	SEAL-ONCE MARINE Penetrating Wood Sealer, Waterproofer & Stain (1 Gallon). Part Number SO7614
		ACE Hardware	Rust-Oleum Marine Coatings Outdoor Gloss White Marine Topside Paint 1 qt. Item no.16754
		Cabela's World's Foremost Outfitter	Parker Coatings Gallon Duck Boat Paint Item: IK-423722
		Lowe's	Rust-Oleum RockSolid Tintable Resurfacer (Actual Net Contents: 116-fl oz) Item # 1030528
		Amazon	

(continued on next page)

Table A3 (continued)

Product	Value Range (USD/kg)	Source	Item
Pesticides	11.08–17.10	Jamestown Distributors	Penofin F3emaga Marine Oil Transparent Oil-based Wood Finish, 1 Gallon Part Number F3EMAGA
		Amazon	TotalBoat TableTop Epoxy
		Wholesale Marine	Waterlox Original Marine Sealer- Gallon by Waterlox Part Number TB 3809 Gallon
		Wholesale Marine	Pettit Flagship Varnish SKU:PET-2015 G.1
		Westmarine	Interlux Schooner Gold Marine Varnish QT SKU:ILX-YVA-500Q
		Jamestown Distributors	PETTTT PAINT–SeaGold Marine Wood Treatment, Quart Model # 17977778
		Wholesale Marine	TotalBoat Lust Marine Varnish
		DoMyOwn	Interlux Sikkens Cetol Marine Wood Finish SKU:ILX-IVA-300.1
		Mosquito Controls	Garden Tech Sevin Insect Killer Concentrate UPC 613499010148
		Mosquito Controls	Malathion 5EC Insecticide, Drexel
		Mosquito Controls	Malathion 57EC, FMC Corporation
		Solutions Pest & Lawn	Malathion 5 EC Insecticide, Winfield
		Do It Yourself	Imidacloprid 2 F Insecticide QUALI-PRO
		DoMyOwn	Permethrin SFR Insecticide 36.8 % Control Solutions
Soil amendments	0.24- 1.72	Amazon	Monterey B.t. Insecticide UPC 022179103900
		eBay	Aries Green Biochar Soil Amendment 5-Gallon Plastic Bucket – USDA, IBI Certified – 100 % Biochar ADIN B07V9VT3MK
		Oregon Biochar Solutions	Rogue Valley Premium Biochar (10 Cu. Yds) Item Number 322550315092
		Pricing	
		Oregon Biochar Solutions	
		Pricing	
Essential oils	11.45-18.72	eBay	Biochar produced from sustainable biomass (ARTIchar) - 55 gallons item number:272304233958
		East Coast Hardware	Premier 0128 P Peat Moss, 2.2 Cu-Ft Bale SKU# 5100155
		BioBizz	BioBizz - Light-Mix 50 Liter Bag (60/Plt) SKU: 722997
			“U.S. Essential Oil Market Size, Share: Industry Analysis Report, 2024.” <i>U.S. Essential Oil Market Size, Share   Industry Analysis Report, 2024</i> , Grand View Research, Feb. 2019, <a href="http://www.grandviewresearch.com/industry-analysis/us-essentia-l-oil-market">www.grandviewresearch.com/industry-analysis/us-essentia-l-oil-market</a> .
			Duncan, Eric. “Topic: Essential Oils.” <i>Www.statista.com</i> , M. Shahbandeh, Aug. 2019, <a href="http://www.statista.com/topics/5174/essential-oils/">www.statista.com/topics/5174/essential-oils/</a> .
			“Essential Oils Market Size, Growth, Share: Global Report 2026.” <i>Essential Oils Market Size, Growth, Share   Global Report 2026</i> , Market Research Report, July 2019, <a href="http://www.fortunebusinessinsights.com/industry-reports/essential-oils-market-101063">www.fortunebusinessinsights.com/industry-reports/essential-oils-market-101063</a> .

Table A4 Product Market Size Data.

Product	Market Size	Reference
Asphalt Binder	IBIS World Market Report	<a href="https://clients1.ibisworld.com/reports/us/industry/majorcompanies.aspx?entid=450">https://clients1.ibisworld.com/reports/us/industry/majorcompanies.aspx?entid=450</a>
	National Asphalt Pavement Association	<a href="http://www.asphaltpavement.org/index.php?option=com_content&amp;view=article&amp;id=891">http://www.asphaltpavement.org/index.php?option=com_content&amp;view=article&amp;id=891</a>
	AsphaltRoad.org	<a href="http://www.asphaltroads.org/assets/_control/content/files/Asphalt_White_Paper_doc.pdf">http://www.asphaltroads.org/assets/_control/content/files/Asphalt_White_Paper_doc.pdf</a>
	National Asphalt Pavement Association	<a href="http://www.asphaltpavement.org/index.php?option=com_content&amp;view=article&amp;id=14&amp;Itemid=33">http://www.asphaltpavement.org/index.php?option=com_content&amp;view=article&amp;id=14&amp;Itemid=33</a>
Adhesives	IBIS World Market Report	<a href="https://clients1.ibisworld.com/reports/us/industry/productsandmarkets.aspx?entid=493">https://clients1.ibisworld.com/reports/us/industry/productsandmarkets.aspx?entid=493</a>
	Freedonia Group	<a href="https://www.freedoniagroup.com/industry-study/adhesives-sealants-3257.htm">https://www.freedoniagroup.com/industry-study/adhesives-sealants-3257.htm</a>
	Market Watch	<a href="https://www.marketwatch.com/press-release/adhesives-and-sealants-market-opportunity-2018-2026-detailed-in-new-research-report-2019-01-24">https://www.marketwatch.com/press-release/adhesives-and-sealants-market-opportunity-2018-2026-detailed-in-new-research-report-2019-01-24</a>
Solventless (laminating) adhesives	GlobeNewswire Reports and Data	<a href="https://www.globenewswire.com/news-release/2019/08/13/1901231/0/en/Adhesives-and-Sealants-Market-To-Rach-USD-88-25-Billion-By-2026-Reports-And-Data.html">https://www.globenewswire.com/news-release/2019/08/13/1901231/0/en/Adhesives-and-Sealants-Market-To-Rach-USD-88-25-Billion-By-2026-Reports-And-Data.html</a>
	Grand View Research	<a href="https://www.grandviewresearch.com/industry-analysis/adhesives-and-sealants-market">https://www.grandviewresearch.com/industry-analysis/adhesives-and-sealants-market</a>
	PR Newswire	<a href="https://www.prnewswire.com/news-releases/the-laminating-adhesives-market-is-projected-to-register-a-cagr-of-6-9-in-terms-of-value-between-2019-and-2024-300941835.html">https://www.prnewswire.com/news-releases/the-laminating-adhesives-market-is-projected-to-register-a-cagr-of-6-9-in-terms-of-value-between-2019-and-2024-300941835.html</a>
Medical Spray Adhesives	PR Newswire	<a href="https://www.prnewswire.com/news-releases/the-laminating-adhesives-market-is-projected-to-register-a-cagr-of-6-9-in-terms-of-value-between-2019-and-2024-300941835.html">https://www.prnewswire.com/news-releases/the-laminating-adhesives-market-is-projected-to-register-a-cagr-of-6-9-in-terms-of-value-between-2019-and-2024-300941835.html</a>
	Markets and Markets	<a href="https://www.marketsandmarkets.com/Market-Reports/spray-adhesive-market-201133268.html">https://www.marketsandmarkets.com/Market-Reports/spray-adhesive-market-201133268.html</a>
Rubber Tackifying resin	Market Watch	<a href="https://www.marketwatch.com/press-release/global-tackifiers-market-size-is-projected-to-be-around-us-4760-milli-on-by-2024-2019-04-12">https://www.marketwatch.com/press-release/global-tackifiers-market-size-is-projected-to-be-around-us-4760-milli-on-by-2024-2019-04-12</a>
	Maximiz Market Research	<a href="https://www.maximizemarketresearch.com/market-report/global-tackifier-market/27433/">https://www.maximizemarketresearch.com/market-report/global-tackifier-market/27433/</a>
Paints and coatings	Ink Wood Research	<a href="https://www.inkwoodresearch.com/reports/north-america-tackifiers-market-forecast-2016-2024/">https://www.inkwoodresearch.com/reports/north-america-tackifiers-market-forecast-2016-2024/</a>
	Statista	<a href="https://www.statista.com/statistics/684695/united-states-paint-and-coatings-demand-by-market/">https://www.statista.com/statistics/684695/united-states-paint-and-coatings-demand-by-market/</a>
	Grand View Research	<a href="https://www.grandviewresearch.com/industry-analysis/paints-coatings-market">https://www.grandviewresearch.com/industry-analysis/paints-coatings-market</a>
Marine coatings	Industry Arc	<a href="https://www.industryarc.com/Report/11726/marine-coatings-market.html">https://www.industryarc.com/Report/11726/marine-coatings-market.html</a>
	Coatings World	<a href="https://www.coatingsworld.com/issues/2018-08-01/view_features/marine-coatings-market-145178/">https://www.coatingsworld.com/issues/2018-08-01/view_features/marine-coatings-market-145178/</a>
	Coatings World	<a href="https://www.coatingsworld.com/issues/2018-08-01/view_features/marine-coatings-market-145178/">https://www.coatingsworld.com/issues/2018-08-01/view_features/marine-coatings-market-145178/</a>
	PR Newswire	

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Table A4 (continued)

Product	Market Size	Reference
Wood preservative coatings	Statista	<a href="https://www.prnewswire.com/news-releases/marine-coatings-market-to-burgeen-at-6-1-cagr-thanks-to-rise-in-shipping-building-activities—tmr-300860524.html">https://www.prnewswire.com/news-releases/marine-coatings-market-to-burgeen-at-6-1-cagr-thanks-to-rise-in-shipping-building-activities—tmr-300860524.html</a>
	Market Watch	<a href="https://www.statista.com/statistics/1062762/global-marine-coating-market-value/">https://www.statista.com/statistics/1062762/global-marine-coating-market-value/</a>
	Coatings World	<a href="https://www.marketwatch.com/press-release/wood-coatings-market-2019-global-industry-share-demand-top-players-industry-size-future-growth-by-2023-market-reports-world-2019-05-09">https://www.marketwatch.com/press-release/wood-coatings-market-2019-global-industry-share-demand-top-players-industry-size-future-growth-by-2023-market-reports-world-2019-05-09</a>
	Tec Navio	<a href="https://www.coatingsworld.com/issues/2016-02-01/view_features/wood-coatings-market-757548/">https://www.coatingsworld.com/issues/2016-02-01/view_features/wood-coatings-market-757548/</a>
Antifouling Paints	Kake News	<a href="https://www.technavio.com/report/global-wood-coatings-market-analysis-share-2018">https://www.technavio.com/report/global-wood-coatings-market-analysis-share-2018</a>
	PR Newswire	<a href="http://www.kake.com/story/41139819/antifouling-coating-market-share-size-2019-industry-growth-factors-top-leaders-development-strategy-future-trends-historical-analysis-competitive">http://www.kake.com/story/41139819/antifouling-coating-market-share-size-2019-industry-growth-factors-top-leaders-development-strategy-future-trends-historical-analysis-competitive</a>
Pesticides	Grand View Research	<a href="https://www.prnewswire.com/news-releases/marine-anti-fouling-coatings-worldwide-market-insights-2019—hull-coatings-segment-to-dominate-the-market-300883344.html">https://www.prnewswire.com/news-releases/marine-anti-fouling-coatings-worldwide-market-insights-2019—hull-coatings-segment-to-dominate-the-market-300883344.html</a>
	IBIS World Market Report	<a href="https://www.grandviewresearch.com/industry-analysis/antifouling-coating-market">https://www.grandviewresearch.com/industry-analysis/antifouling-coating-market</a>
	Ag Professional	<a href="https://clients1.ibisworld.com/reports/us/industry/productsandmarkets.aspx?entid=484">https://clients1.ibisworld.com/reports/us/industry/productsandmarkets.aspx?entid=484</a>
	Pesticide Action Network North America	<a href="https://www.agprofessional.com/article/study-shows-global-pesticide-market-reach-81-billion-five-years">https://www.agprofessional.com/article/study-shows-global-pesticide-market-reach-81-billion-five-years</a>
Essential Oils	Grand View Research	<a href="http://www.panna.org/blog/us-and-world-pesticide-use">http://www.panna.org/blog/us-and-world-pesticide-use</a>
	Statista	<a href="https://www.panna.org/blog/us-and-world-pesticide-use">https://www.panna.org/blog/us-and-world-pesticide-use</a>
	Fortune Business Insight	<a href="https://www.grandviewresearch.com/industry-analysis/essential-oils-market">https://www.grandviewresearch.com/industry-analysis/essential-oils-market</a>
	IBIS World Market Report	<a href="https://www.statista.com/topics/5174/essential-oils/">https://www.statista.com/topics/5174/essential-oils/</a>
		<a href="https://www.fortunebusinessinsights.com/industry-reports/essential-oils-market-101063">https://www.fortunebusinessinsights.com/industry-reports/essential-oils-market-101063</a>
		<a href="https://clients1.ibisworld.com/reports/us/industry/productsandmarkets.aspx?entid=5061">https://clients1.ibisworld.com/reports/us/industry/productsandmarkets.aspx?entid=5061</a>

Table A5

Typical fractions of active ingredients contained in pesticide products.

Product Category	Active Ingredient Percentage	Source	Item
Termiticide, Insecticide and Fungicide	40 %	Forestry Distributing, North America's Forestry Leader	Niscus Corporation Bora-Care Termiticide, Insecticide & Fungicide, Nisus
Wood Preservative coatings	25 %	Perma-chink Systems, Inc.	Shell Guard Concentrate Borate Glycol Wood Preservative Coating
Termiticide and Insecticide	36 %	DoMyOwn	Tengard SFR Termiticide Insecticide
Insecticide	21 %	Solutions Pest & Lawn	CSI Dominion 2 I Insecticide
Termiticide and Insecticide	9 %	Do-It-Yourself Pest Control	Control Solutions Taurus SC Insecticide-Termiticide

## References

- Abdel-Haleem, H., Foster, M., Ray, D., Coffelt, T., 2018. Phenotypic variations, heritability and correlations in dry biomass, rubber and resin production among guayule improved germplasm lines. *Ind. Crops Prod.* 112, 691–697. <https://doi.org/10.1016/j.indcrop.2017.12.072>.
- Alcántara-Flores, E., Brechú-Franco, E.A., García-López, P., Rocha-Zavaleta, L., López-Marure, R., Martínez-Vázquez, M., 2015. Argentatin B inhibits proliferation of prostate and colon cancer cells by inducing cell senescence. *Molecules* 20. <https://doi.org/10.3390/molecules201219757>.
- Angulo-Sánchez, J.L., Neira-Velázquez, G., de Rodríguez, D.J., 1995. Multimodal molecular weight distributions of guayule rubber from high rubber yielding lines. *Ind. Crops Prod.* 4, 113–120. [https://doi.org/10.1016/0926-6690\(95\)00023-6](https://doi.org/10.1016/0926-6690(95)00023-6).
- Armistead, J.P., Bultman, J.D., Griffith, J.R., 1987. Guayule resin as an extender for an epoxy system. Seventh Annual Conference of the Guayule Rubber Society, Annapolis, Maryland 2–6.
- Awouafack, M.D., Tane, P., Kuete, V., Eloff, J.N., 2013. 2 - Sesquiterpenes from the medicinal plants of Africa. In: Kuete, V. (Ed.), *Medicinal Plant Research in Africa*. Elsevier, Oxford, pp. 33–103.
- Backhaus, R.A., Walsh, S., 1983. The ontogeny of rubber formation in guayule, *Parthenium argentatum* Gray. *Bot. Gaz.* 144, 391–400. <https://doi.org/10.1086/337388>.
- Bajwa, D.S., Holt, G.A., Bajwa, S.G., Duke, S.E., McIntyre, G., 2017. Enhancement of termite (*Reticulitermes flavipes* L.) resistance in mycelium reinforced biofiber-composites. *Ind. Crops Prod.* 107, 420–426. <https://doi.org/10.1016/j.indcrop.2017.06.032>.
- Banigan, T.F., Meeks, J.W., 1953. Isolation of palmitic, stearic and linoleic acids from guayule resin1. *J. Am. Chem. Soc.* 75, 3829–3830. <https://doi.org/10.1021/ja01111a501>.
- Banigan Jr., T.F., Meeks, J.W., Planck, R.W., 1951. Distribution of waxes in guayule. *Bot. Gaz.* 113, 231–234. <https://doi.org/10.1086/335716>.
- Banigan, T.F., Verbiscar, A.J., Weber, C.W., 1982. Composition of guayule leaves, seed and wood. *J. Agric. Food Chem.* 30, 427–431. <https://doi.org/10.1021/jf00111a004>.
- Battistel, E., Querci, C., Ramello, S., 2017. Integrated process for processing and utilising the guayule plant. Versalis S.p.A. US Patent 9969818 B2.
- Beinor, R.T., Cole, W.M., 1986. Solvent fractionation of guayule rubber. *Bridgestone Firestone Inc. US Patent* 4623713 A.
- Belmares, H., Jimenez, L.L., Ortega, M., 1980. New rubber peptizers and coatings derived from guayule resin (*Parthenium argentatum* Gray). *Ind. Eng. Chem. Res.* 19, 107–111. <https://doi.org/10.1021/i360073a025>.
- Black, L.T., Hamerstrand, G.E., Nakayama, F.S., Rasnik, B.A., 1983. Gravimetric analysis for determining the resin and rubber content of guayule. *Rubber Chem Technol* 56, 367–371. <https://doi.org/10.1021/i360073a025>.
- Black, L.T., Swanson, C.L., Hamerstrand, G.E., 1986. Effects of storage on the molecular weight of rubber contained in guayule. *Rubber Chem Technol* 59, 123–129. <https://doi.org/10.5254/1.3538179>.
- Boateng, A.A., Mullen, C.A., McMahan, C.M., Whalen, M.C., Cornish, K., 2010. Guayule (*Parthenium argentatum*) pyrolysis and analysis by PY-GC/MS. *Anal Appl Pyrolysis* 87, 14–23. <https://doi.org/10.1016/j.jaap.2009.09.005>.
- Bonner, J., 1991. The history of rubber. In: Whitworth, J.W., Whitehead, E.E. (Eds.), *Guayule Natural Rubber: a Technical Publication With Emphasis on Recent Findings*. GAMC-USDA/CSRS, Tucson, AZ, pp. 1–16.
- Buchanan, R.A., Cull, I.M., Otey, F.H., Russell, C.R., 1978. Hydrocarbon-and rubber-producing crops. *Econ. Bot.* 32, 146–153. <https://doi.org/10.1007/BF02866868>.
- Buchanan, B.B., Gruissem, W., Jones, R.L., 2015. *Biochemistry and Molecular Biology of Plants*. John Wiley & Sons.
- Bultman, J.D., Beal, R.H., Bailey, C.A., Schloman, W.W., 1986. The evaluation of non-rubber extractives from the guayule plant (*Parthenium argentatum* Gray) for pesticidal worth. Seventeenth Annual Meeting of the International Research Group on Wood Preservation.
- Bultman, J.D., Gilbertson, R.L., Adaskaveg, J., Amburgey, T.L., Parikh, S.V., Bailey, C.A., 1991. The efficacy of guayule resin as a pesticide. *Bioresour. Technol.* 35, 197–201. [https://doi.org/10.1016/0960-8524\(91\)90030-N](https://doi.org/10.1016/0960-8524(91)90030-N).
- Bultman, J.D., Chen, S.-L., Schloman, W.W., 1998. Anti-termite efficacy of the resin and rubber in fractionator overheads from a guayule extraction process. *Ind. Crops Prod.* 8, 133–143. [https://doi.org/10.1016/S0926-6690\(97\)10018-8](https://doi.org/10.1016/S0926-6690(97)10018-8).
- Campos-López, E., Angulo-Sánchez, J.L., 1976. Molecular weight characterization of natural guayule rubber by gel-permeation chromatography. *J. Polym. Sci. B* 14, 649–652. <https://doi.org/10.1002/pol.1976.130141103>.
- Carlos, L.C., Mariano, M.-V., Jose, S.C., Juan, R.S., Eduardo, A., 2001. Insect growth regulatory activity of some extracts and compounds from *Parthenium argentatum* on fall armyworm *Spodoptera frugiperda*. *Z Naturforsch C J Biosci.* 56, 95–105. <https://doi.org/10.1515/znc-2001-1-216>.
- Cheng, F., Dehghanizadeh, M., Audu, M.A., Jarvis, J.M., Holguin, F.O., Brewer, C.E., 2020. Characterization and evaluation of guayule processing residues as potential feedstock for biofuel and chemical production. *Ind. Crops Prod.* 150, 112311. <https://doi.org/10.1016/j.indcrop.2020.112311>.
- Chow, P., Nakayama, F.S., Youngquist, J.A., Muehl, J.H., Krzysik, A.M., 2002. Durability of wood/plastic composites made from *Parthenium* species. In: Thirty-Third Annual Meeting of the International Research Group on Wood Preservation, Section 4, Processes and Properties. Stockholm, Sweden: IRG Secretariat, 2002. Cardiff, Wales, p. 12.
- Cole, W.M., Hilton, A.S., Schloman Jr., W.W., Compton, J.B., Dembek Jr., J.A., 1991. *Guayule Rubber Project: Final Report Prepared Under Contract No 53-3142-7-6005*. Bridgestone/Firestone, Akron, Ohio.
- Cornish, K., 1996. Hypoallergenic natural rubber products from *Parthenium argentatum* (gray) and other non-hevea brasiliensis species. United States Dept. of Agriculture. US Patent 5580942 A.

- Cornish, K., 2017. Alternative natural rubber crops: why should we care? *Technol. Innov.* 18, 244–255. <https://doi.org/10.21300/18.4.2017.245>.
- Cornish, K., Backhaus, R.A., 1990. Rubber transferase activity in rubber particles of guayule. *Phytochemistry* 29, 3809–3813. [https://doi.org/10.1016/0031-9422\(90\)85337-F](https://doi.org/10.1016/0031-9422(90)85337-F).
- Cornish, K., Martin, J.A., Marentis, R.T., Plamthottam, S., 2006. Extraction and fractionation of biopolymers and resins from plant materials. Yulex Corporation. US Patent 7259231 B2.
- Cornish, K., Pearson, C.H., Rath, D.J., 2013. Accurate quantification of guayule resin and rubber requires sample drying below a critical temperature threshold. *Ind. Crops Prod.* 41, 158–164. <https://doi.org/10.1016/j.indcrop.2012.04.014>.
- Cornish, K., Marentis, R.T., 2010. Rapid expanded solvent extraction. Yulex Corporation. US Patent 7790036 B2.
- Costa, E., Aguado, J., Ovejero, G., Cañizares, P., 1992. Conversion of guayule resin to C1-C10 hydrocarbons on zeolite based catalysts. *Fuel* 71, 109–113. [https://doi.org/10.1016/0016-2361\(92\)90200-8](https://doi.org/10.1016/0016-2361(92)90200-8).
- Curtis Jr., O.F., 1947. Distribution of rubber and resins in guayule. *Plant Physiol.* 22, 333–359. <https://doi.org/10.1104/pp.22.4.333>.
- D.S. Winkler, H.L.S., 1977. Plasticification effect of guayule resin on raw rubber. In: *The 2nd International Guayule Conference*. Salti Ilo, Mexico.
- Daly, M.A., 1989. Characterization and Fractionation by Ultrafiltration of Guayule Resin, Chemical Engineering. Texas A&M University. <https://hdl.handle.net/1969.1/etd-tamu-1989-thesis-d1535>.
- de Vivar, A.R., Martínez-Vázquez, M., Matsubara, C., Pérez-Sánchez, G., Joseph-Nathan, P., 1990. Triterpenes in *Parthenium argentatum*, structures of argentinins C and D. *Phytochemistry* 29, 915–918. [https://doi.org/10.1016/0031-9422\(90\)80045-1](https://doi.org/10.1016/0031-9422(90)80045-1).
- Dehghanizadeh, M., Cheng, F., Jarvis, J.M., Holguin, F.O., Brewer, C.E., 2020. Characterization of resin extracted from guayule (*Parthenium argentatum*): a dataset including GC-MS and FT-ICR MS. *Data Brief* 105989. <https://doi.org/10.1016/j.dib.2020.105989>.
- Díaz-Maroto, M.C., Pérez-Coello, M.S., González Viñas, M.A., Cabezedo, M.D., 2003. Influence of drying on the flavor quality of spearmint (*Mentha spicata* L.). *J. Agric. Food Chem.* 51, 1265–1269. <https://doi.org/10.1021/jf020805l>.
- Dong, J., Ma, X., Fu, Z., Guo, Y., 2011. Effects of microwave drying on the contents of functional constituents of *Eucommia ulmoides* flower tea. *Ind. Crops Prod.* 34, 1102–1110. <https://doi.org/10.1016/j.indcrop.2011.03.026>.
- Estilai, A., 1991. Biomass, rubber, and resin yield potentials of new guayule germplasm. *Bioresour. Technol.* 35, 119–125. [https://doi.org/10.1016/0960-8524\(91\)90018-F](https://doi.org/10.1016/0960-8524(91)90018-F).
- Finlay, M.R., 2009. *Growing American Rubber: Strategic Plants and the Politics of National Security*. Rutgers University Press.
- Foster, M.A., Coffelt, T.A., 2005. Guayule agronomics: establishment, irrigated production, and weed control. *Ind. Crops Prod.* 22, 27–40. <https://doi.org/10.1016/j.indcrop.2004.06.006>.
- Gaytán, P., Yañez, J., Soberón, X., Martínez, R., 1997. A new method for oligonucleotide derivatization of the 3' or 5' termini with a CPG-support carrying the natural product isogentatin-D. *Tetrahedron Lett.* 38, 6123–6126. [https://doi.org/10.1016/S0040-4039\(97\)01388-9](https://doi.org/10.1016/S0040-4039(97)01388-9).
- Ghasemi Pirbalouti, A., Mahdad, E., Craker, L., 2013. Effects of drying methods on qualitative and quantitative properties of essential oil of two basil landraces. *Food Chem.* 141, 2440–2449. <https://doi.org/10.1016/j.foodchem.2013.05.098>.
- Gilliland, M.G., van Staden, J., 1986. Cyclic patterns of growth and rubber deposition in guayule *Parthenium argentatum*. Suggestions for a management programme. *S. Afr. J. Plant Soil* 3, 21–26. <https://doi.org/10.1080/02571862.1986.10634180>.
- Gilliland, M.G., Appleton, M.R., Van Staden, J., 1988. Gland cells in resin canal epithelia in guayule (*Parthenium argentatum*) in relation to resin and rubber production. *Ann. Bot.* 61, 55–64. <https://doi.org/10.1093/oxfordjournals.aob.a087527>.
- Gumbs, R.W., 2004. *Low Cost Solventless Guayule Resin Adhesives. Final Report to US Department of Agriculture on Grant No. 2004-33610-14337*, pp. 1–34.
- Gumbs, R.W., 2008. Guayule rubber-containing wet-stick bioadhesives. Yulex Corporation. US Patent 20080300526 A1.
- Gumbs, R.W., 2009. Guayule resin multipolymers. Yulex Corporation. US Patent 20090099309 A1.
- Gutiérrez, C., Gonzalez-Coloma, A., Hoffmann, J.J., 1999. Antifeedant properties of natural products from *Parthenium argentatum* x *P. tomentosum* (Asteraceae) and *Castela emoryi* (Simaroubaceae) against *Reticulitermes flavipes*. *Ind. Crops Prod.* 10, 35–40. [https://doi.org/10.1016/S0926-6690\(99\)00003-5](https://doi.org/10.1016/S0926-6690(99)00003-5).
- Gutierrez, R., Kay, E.L., 1983. Processing of guayule material by volatilizing and heating steps. Bridgestone Firestone Inc. US Patent 4376853 A.
- Haagen-Smit, A.J., Siu, R., 1944. *Chemical Investigations in Guayule. I. Essential Oil of Guayule, Parthenium argentatum, Gray*. *J. Am. Chem. Soc.* 66, 2068–2074. <https://doi.org/10.1021/ja01240a019>.
- Hamrouni-Sellami, I., Rahali, F.Z., Rebey, I.B., Bourgo, S., Limam, F., Marzouk, B., 2013. Total phenolics, flavonoids, and antioxidant activity of sage (*Salvia officinalis* L.) plants as affected by different drying methods. *Food Bioproc. Tech.* 6, 806–817. <https://doi.org/10.1007/s11947-012-0877-7>.
- Hemida, A., Abdelrahman, M., 2019. Influence of guayule resin as a bio-based additive on asphalt-rubber binder at elevated temperatures. *Recycling* 4. <https://doi.org/10.3390/recycling4030038>.
- Huang, H.T., 1991. Introduction. In: Whitworth, J.W., Whitehead, E.E. (Eds.), *Guayule Natural Rubber: a Technical Publication With Emphasis on Recent Findings*. GAMC-USDA/CSRS, Tucson, AZ, pp. xv–xix.
- Isman, M.B., Rodriguez, E., 1983. Larval growth inhibitors from species of *Parthenium* (Asteraceae). *Phytochemistry* 22, 2709–2713. [https://doi.org/10.1016/S0031-9422\(00\)97677-5](https://doi.org/10.1016/S0031-9422(00)97677-5).
- Jeyaseelan, R.S., Wagner, J.P., 1995. Ultrafiltration of guayule resin. *J. Memb. Sci.* 103, 45–50. [https://doi.org/10.1016/0376-7388\(94\)00305-1](https://doi.org/10.1016/0376-7388(94)00305-1).
- Jones, E.P., 1948. Recovery of rubber latex from Guayule Shrub. *Ind. Eng. Chem. Res.* 40, 864–874. <https://doi.org/10.1021/ie50461a019>.
- Joseph, J.P., Shah, J.J., Inamdar, J.A., 1988. Distribution, development and structure of resin ducts in guayule (*Parthenium argentatum* Gray). *Ann. Bot.* 61, 377–387. <https://doi.org/10.1093/oxfordjournals.aob.a087567>.
- Kay, E.L., Gutierrez, R., 1985. *Rubber Additives Derived From Guayule Resins and Compositions Containing Them*. Bridgestone Firestone Inc. 4542191 A.
- Kay, E.L., Gutierrez, R., 1984. Process for extracting rubber and by-products from guayule and guayule-like shrubs. Bridgestone Firestone Inc. US Patent 4435337 A.
- Keller, R.W., Winkler, D.S., Stephens, H.L., 1981. Degradative effects of guayule resin on natural rubber. *Rubber Chem Technol* 54, 115–123. <https://doi.org/10.5254/1.3535786>.
- Lloyd, F.E., 1911. *Guayule (Parthenium Argentatum Gray): a Rubber-plant of the Chihuahuan Desert*. Carnegie Institution of Washington.
- Lusher, S.M., Richardson, D.N., 2015. Guayule plant extracts as recycling agents in hot mix asphalt with high reclaimed binder content. *J. Mater. Civ. Eng.* 27. [https://doi.org/10.1061/\(ASCE\)MT.1943-5533.0001238](https://doi.org/10.1061/(ASCE)MT.1943-5533.0001238), 04014269-1-04014269-10.
- Maatooq, G.T., Hoffmann, J.J., 1996. Fungistatic sesquiterpenoids from *Parthenium*. *Phytochemistry* 43, 67–69. [https://doi.org/10.1016/0031-9422\(96\)00225-7](https://doi.org/10.1016/0031-9422(96)00225-7).
- Maatooq, G.T., Hoffmann, J.J., 2002. Microbial transformation of a mixture of Argentinin A and Incanilin. *Z. Naturforsch. C J. Biosci.* 57, 489–495. <https://doi.org/10.1515/znc-2002-5-615>.
- Maatooq, G.T., Stumpf, D.K., Hoffmann, J.J., Hutter, L.K., Timmermann, B.N., 1996. Antifungal eudesmanoids from *Parthenium argentatum* x *P. tomentosum*. *Phytochemistry* 41, 519–524. [https://doi.org/10.1016/0031-9422\(95\)00580-3](https://doi.org/10.1016/0031-9422(95)00580-3).
- Mabry, T.J., Hunziker, J.H., Difeo Jr., D.R., 1978. *Cresosote Bush: Biology and Chemistry of Larrea in New World Deserts*. Dowden, Hutchinson & Ross, Inc.
- Madhavan, S., Benedict, C.R., 1984. Isopentenyl pyrophosphate cis-1,4-Polyisoprenyl Transferase from guayule (*Parthenium argentatum* Gray). *Plant Physiol.* 75, 908. <https://doi.org/10.1104/pp.75.4.908>.
- Martínez, M., Flores, G., de Vivar, A.R., Reynolds, G., Rodriguez, E., 1986. Guayulins C and d from guayule (*Parthenium argentatum*). *J. Nat. Prod.* 49, 1102–1103. <https://doi.org/10.1021/np50048a022>.
- Martínez, R., Martínez-Vázquez, M., Zamorano-Macías, O., 1990. Electron impact mass spectrometry of triterpenoids from guayule. *Org. Mass Spectrom* 25, 237–238. <https://doi.org/10.1002/oms.1210250411>.
- Martínez Vazquez, M., Martínez, R., Espinosa Perez, G., Díaz, M., Herrera Sanchez, M., 1994. Antimicrobial properties of argentinin A, isolated from *Parthenium argentatum*. *Fitoterapia* 65, 371–372.
- Mears, J.A., 1980. The flavonoids of *Parthenium* I. *J. Nat. Prod.* 43, 708–716. <https://doi.org/10.1021/np50012a002>.
- Monadjemi, S.M.A., McMahan, C.M., Cornish, K., 2016. Effect of non-rubber constituents on guayule and Hevea rubber intrinsic properties. *J. Res. Updates Polym. Sci.* 5, 87–96. <https://doi.org/10.6000/1929-5995.2016.05.03.1>.
- Mooibroek, H., Cornish, K., 2000. Alternative sources of natural rubber. *Appl. Microbiol. Biotechnol.* 53, 355–365. <https://doi.org/10.1007/s002530051627>.
- Nakayama, F.S., 2005. Guayule future development. *Ind. Crops Prod.* 22, 3–13. <https://doi.org/10.1016/j.indcrop.2004.05.006>.
- Nakayama, F.S., Vinyard, S.H., Chow, P., Bajwa, D.S., Youngquist, J.A., Muehl, J.H., Krzysik, A.M., 2001. Guayule as a wood preservative. *Ind. Crops Prod.* 14, 105–111. [https://doi.org/10.1016/S0926-6690\(00\)00093-5](https://doi.org/10.1016/S0926-6690(00)00093-5).
- Nakayama, F.S., Chow, P., Vinyard, S.H., Depe, N.A., Faber, A.L., 2003. Termite resistance of kenaf composite boards treated with guayule resin. *Association for the Advancement of Industrial Crops Annual Meeting Program and Abstracts, Portland, Oregon* 61.
- Nakayama, F.S., Youngquist, J.A., Chow, P., 2004. Composites comprising plant material from *Parthenium* spp. and plastic. United States Dept. of Agriculture. US Patent 6787590 B2.
- Ownby, D.R., Ownby, H.E., McCullough, J., Shafer, A.W., 1996. The prevalence of anti-latex IgE antibodies in 1000 volunteer blood donors. *J. Allergy Clin. Immunol.* 97, 1188–1192. [https://doi.org/10.1016/S0091-6749\(96\)70183-5](https://doi.org/10.1016/S0091-6749(96)70183-5).
- Parodi, F.J., 1988. *Structure Elucidation of Natural Products From Asteraceae by Modern NMR Techniques and Biomimetic Transformations of 11,13-dihydroparthenolide*. Louisiana State University. [https://digitalcommons.lsu.edu/gradschool\\_disstheses/4528](https://digitalcommons.lsu.edu/gradschool_disstheses/4528).
- Parra-Delgado, H., García-Pillado, F., Sordo, M., Ramírez-Apan, T., Martínez-Vázquez, M., Ostrosky-Wegman, P., 2005. Evaluation of the cytotoxicity, cytostaticity and genotoxicity of Argentinins A and B from *Parthenium argentatum* (Gray). *Life Sci.* 77, 2855–2865. <https://doi.org/10.1016/j.lfs.2005.05.024>.
- Pascual-Villalobos, M.J., López, M.D., 2013. New application of guayule resin in controlled release formulations. *Ind. Crops Prod.* 43, 44–49. <https://doi.org/10.1016/j.indcrop.2012.07.001>.
- Pearson, C.H., Cornish, K., Rath, D.J., 2013. Extraction of natural rubber and resin from guayule using an accelerated solvent extractor. *Ind. Crops Prod.* 43, 506–510. <https://doi.org/10.1016/j.indcrop.2012.06.052>.
- Piana, F., Ciulu, M., Quirantes-Piné, R., Sanna, G., Segura-Carretero, A., Spano, N., Mariani, A., 2018. Simple and rapid procedures for the extraction of bioactive compounds from Guayule leaves. *Ind. Crops Prod.* 116, 162–169. <https://doi.org/10.1016/j.indcrop.2018.02.057>.
- Piluzza, G., Campesi, G., Molinu, M.G., Re, G.A., Sulas, L., 2020. Bioactive compounds from leaves and twigs of guayule grown in a Mediterranean environment. *Plants* 9. <https://doi.org/10.3390/plants9040442>.

- Punvichai, T., Amor, A., Tardan, E., Palu, S., Pioch, D., 2016. SC-CO<sub>2</sub> Extraction of guayule biomass (*Parthenium argentatum*) - yield and selectivity towards valuable co-products, lipids and terpenics. *Biointerface Res Appl Chem* 6, 1777–1787.
- Randall, A.M., Calliano, L., Galizio, B.C., Di Ronza, R., Aurisicchio, C., Pezzullo, G., 2020. Tire sealant composition. Bridgestone Corporation, Japan; Bridgestone Americas Tire Operations, LLC. US Patent 10675822 B2.
- Ray, D.T., 1993. Guayule: a source of natural rubber. In: Janick, J., Simon, J.E. (Eds.), *New Crops*. John Wiley & Sons Ltd., New York, pp. 338–343.
- Romero-Benavides, J.C., Bailon-Moscoco, N., Parra-Delgado, H., Ramirez, M.I., Villalís, J., Cabrera, H., Gonzalez-Arevalo, G., Cueva, R., Zentella-Dehesa, A., Ratovitski, E.A., Martínez-Vázquez, M., 2018. Argentatin B derivatives induce cell cycle arrest and DNA damage in human colon cancer cells through p73/p53 regulation. *Med. Chem. Res.* 27, 834–843. <https://doi.org/10.1007/s00044-017-2106-4>.
- Rozalén, J., García, M.M., Zalacain, A., López-Córcoles, H., Hurtado de Mendoza, J., Cornish, K., Carmona, M., 2021. Future trends for the analysis of guayulins in guayule (*Parthenium argentatum* Gray) resins. *Ind. Crops Prod.* 159, 113027. <https://doi.org/10.1016/j.indcrop.2020.113027>.
- Sabaini, P.S., Boateng, A.A., Schaffer, M., Mullen, C.A., Elkasabi, Y., McMahan, C.M., Macken, N., 2018. Techno-economic analysis of guayule (*Parthenium argentatum*) pyrolysis biorefining: production of biofuels from guayule bagasse via tail-gas reactive pyrolysis. *Ind. Crops Prod.* 112, 82–89. <https://doi.org/10.1016/j.indcrop.2017.11.009>.
- Saldívar, R.H.L., García, G.F.B., Castillo, F.D.H., Alvarado, R.G., de Rodríguez, D.J., Díaz, F.J., 2003. Evaluation of resin content and the antifungal effect of *Larrea tridentata* (Sesse and Moc. Ex DC) Coville extracts from two Mexican deserts against *Phythium* sp. *Pringsh. Rev. Mex. Fitopatol.* 21, 97–101.
- Salvucci, M.E., Coffelt, T.A., Cornish, K., 2009. Improved methods for extraction and quantification of resin and rubber from guayule. *Ind. Crops Prod.* 30, 9–16. <https://doi.org/10.1016/j.indcrop.2008.12.006>.
- Sarmiento, L.A.V., Spricigo, C.B., Petrus, J.C.C., Carlson, L.H.C., Machado, R.A.F., 2004. Performance of reverse osmosis membranes in the separation of supercritical CO<sub>2</sub> and essential oils. *J. Memb. Sci.* 237, 71–76. <https://doi.org/10.1016/j.memsci.2004.02.021>.
- Sarmiento, L.A.V., Machado, R.A.F., Petrus, J.C.C., Tamanini, T.R., Bolzan, A., 2008. Extraction of polyphenols from cocoa seeds and concentration through polymeric membranes. *J. Supercrit. Fluids* 45, 64–69. <https://doi.org/10.1016/j.supflu.2007.11.007>.
- Schloman, W.W. Jr., 1988a. Process for the controlled partition of guayule resin. Bridgestone Firestone Inc. US Patent 4739038 A.
- Schloman Jr., W.W., 1988b. Rubber additives derived from guayule resin. *Ind. Eng. Chem. Res.* 27, 712–716. <https://doi.org/10.1021/ie00076a031>.
- Schloman, W.W., Jr., 1991. Free-flowing guayule resin and bagasse mixtures and their use as fuel or soil amendent. Bridgestone Firestone Inc. US Patent 4988388 A.
- Schloman, W.W., 1992. Low-molecular-weight guayule natural rubber as a feedstock for epoxidized natural rubber. *Ind. Crops Prod.* 1, 35–39. [https://doi.org/10.1016/0926-6690\(92\)90043-U](https://doi.org/10.1016/0926-6690(92)90043-U).
- Schloman, W.W., 2005. Processing guayule for latex and bulk rubber. *Ind. Crops Prod.* 22, 41–47. <https://doi.org/10.1016/j.indcrop.2004.04.031>.
- Schloman Jr., W.W., Wagner, J.P., 1991. Rubber and coprodut utilization. In: Whitworth, J.W., Whitehead, E.E. (Eds.), *Guayule Natural Rubber: a Technical Publication with Emphasis on Recent Findings*. GAMC-USDA/CSRS, Tucson, AZ, pp. 287–310.
- Schloman, W.W., Hively, R.A., Krishen, A., Andrews, A.M., 1983. Guayule byproduct evaluation: extract characterization. *J. Agric. Food Chem.* 31, 873–876. <https://doi.org/10.1021/jf00118a050>.
- Schloman, W.W., Garrot, D.J., Ray, D.T., Bennett, D.J., 1986. Seasonal effects on guayule resin composition. *J. Agric. Food Chem.* 34, 177–179. <https://doi.org/10.1021/jf00068a005>.
- Schloman, W.W., Carlson, D.W., Hilton, A.S., 1988. Guayule extractables: influence of extraction conditions on yield and composition. *Biomass* 17, 239–249. [https://doi.org/10.1016/0144-4565\(88\)90106-0](https://doi.org/10.1016/0144-4565(88)90106-0).
- Schloman Jr., W.W., Wyzgoski, F., McIntyre, D., Cornish, K., Siler, D.J., 1996. Characterization and performance testing of guayule latex. *Rubber Chem Technol* 69, 215–222. <https://doi.org/10.5254/1.3538366>.
- Schloman, W.W., Davis, J.A., 1986. Sulfurized guayule resin and rubber. Bridgestone Firestone Inc. US Patent 4621118 A.
- Scora, R.W., Kumamoto, J., 1979. Essential leaf oils of *Parthenium argentatum* A. Gray. *J. Agric. Food Chem.* 27, 642–643. <https://doi.org/10.1021/jf60223a016>.
- Sfeir, N., Chapuset, T., Palu, S., Lançon, F., Amor, A., García, J.G., Snoeck, D., 2014. Technical and economic feasibility of a guayule commodity chain in Mediterranean Europe. *Ind. Crops Prod.* 59, 55–62. <https://doi.org/10.1016/j.indcrop.2014.04.043>.
- Shen, M.C., Rodríguez, E., Kerr, K., Mabry, T.J., 1976. Flavonoids of four species of *Parthenium* (compositae). *Phytochemistry* 15, 1045–1047. [https://doi.org/10.1016/S0031-9422\(00\)84401-5](https://doi.org/10.1016/S0031-9422(00)84401-5).
- Sidhu, O.P., Ratti, N., Behl, H.M., 1995. Quantitative and qualitative variations in resin content and guayulins (A and B) among different guayule cultivars. *J. Agric. Food Chem.* 43, 2012–2015. <https://doi.org/10.1021/jf00056a010>.
- Siler, D.J., Cornish, K., Hamilton, R.G., 1996. Absence of cross-reactivity of IgE antibodies from subjects allergic to *Hevea brasiliensis* latex with a new source of natural rubber latex from guayule (*Parthenium argentatum*). *J. Allergy Clin. Immunol.* 98, 895–902. [https://doi.org/10.1016/S0091-6749\(96\)80005-4](https://doi.org/10.1016/S0091-6749(96)80005-4).
- Singh, B.P. (Ed.), 2010. *Industrial Crops and Uses*. CABI, p. 512.
- Smith, L.L., Wagner, J.P., Rock, J.C., 1996. Smoke and toxic species analyses from controlled combustion of wood impregnated with guayule resin. *Polym. Technol. Eng.* 35, 877–904. <https://doi.org/10.1080/03602559608000605>.
- Snoeck, D., Chapuset, T., García García, J., Sfeir, N., Palu, S., 2015. Feasibility of a guayule commodity chain in the Mediterranean region. *Ind. Crops Prod.* 75, 159–164. <https://doi.org/10.1016/j.indcrop.2015.05.008>.
- Spano, N., Meloni, P., Idda, I., Mariani, A., Pilo, I.M., Nurchi, M.V., Lachowicz, L.J., Rivera, E., Orona-Espino, A., Sanna, G., 2018. Assessment, validation and application to real samples of an RP-HPLC method for the determination of guayulins a, B, C and d in Guayule Shrub. *Separations* 5. <https://doi.org/10.3390/separations5020023>.
- Spricigo, C.B., Bolzan, A., Machado, R.A.F., Carlson, L.H.C., Petrus, J.C.C., 2001. Separation of nutmeg essential oil and dense CO<sub>2</sub> with a cellulose acetate reverse osmosis membrane. *J. Memb. Sci.* 188, 173–179. [https://doi.org/10.1016/S0376-7388\(01\)00353-2](https://doi.org/10.1016/S0376-7388(01)00353-2).
- Springob, K., Kutchan, T.M., 2009. *Introduction to the Different Classes of Natural Products, Plant-Derived Natural Products*. Springer, pp. 3–50.
- Sproul, E., Summers, H.M., Seavert, C., Robbs, J., Khanal, S., Mealing, V., Landis, A.E., Fan, N., Sun, O., Quinn, J.C., 2020. Integrated techno-economic and environmental analysis of guayule rubber production. *J. Clean. Prod.* 273, 122811. <https://doi.org/10.1016/j.jclepro.2020.122811>.
- Srinivasan, N., Ju, L.-K., 2010. Pretreatment of guayule biomass using supercritical carbon dioxide-based method. *Bioresour. Technol.* 101, 9785–9791. <https://doi.org/10.1016/j.biortech.2010.07.069>.
- Strickland, B., 2013. Defining and qualifying various types of adhesives. In: *Toxic Use Reduction Institute (TURI) Spring Continuing Education Conference*, Sturbridge Host Hotel and Conference Center. Sturbridge, MA.
- Suchat, S., Pioch, D., Palu, S., Tardan, E., van Loo, E.N., Davrieux, F., 2013. Fast determination of the resin and rubber content in *Parthenium argentatum* biomass using near infrared spectroscopy. *Ind. Crops Prod.* 45, 44–51. <https://doi.org/10.1016/j.indcrop.2012.09.025>.
- Swift, K.A.D., 2004. Catalytic transformations of the major terpene feedstocks. *Top. Catal.* 27, 143–155. <https://doi.org/10.1023/B:TOCA.0000013549.60930.da>.
- Szumny, A., Figiel, A., Gutiérrez-Ortiz, A., Carbonell-Barrachina, Á.A., 2010. Composition of rosemary essential oil (*Rosmarinus officinalis*) as affected by drying method. *J. Food Eng.* 97, 253–260. <https://doi.org/10.1016/j.jfoodeng.2009.10.019>.
- Taurines, M., Brancheriau, L., Palu, S., Pioch, D., Tardan, E., Boutahar, N., Sartre, P., Meunier, F., 2019. Determination of natural rubber and resin content of guayule fresh biomass by near infrared spectroscopy. *Ind. Crops Prod.* 134, 177–184. <https://doi.org/10.1016/j.indcrop.2019.03.073>.
- Tavarez-Santamaría, Z., Jacobo-Herrera, N.J., Rocha-Zavaleta, L., Zentella-Dehesa, A., Couder-García, B.D., Martínez-Vázquez, M., 2020. A higher frequency administration of the nontoxic cycloartane-type triterpene argentatin a improved its anti-tumor activity. *Molecules* 25. <https://doi.org/10.3390/molecules25081780>.
- Teetor, V.H., Ray, D.T., Schloman, W.W., 2009. Evaluating chemical indices of guayule rubber content: guayulins A and B. *Ind. Crops Prod.* 29, 590–598. <https://doi.org/10.1016/j.indcrop.2008.11.005>.
- Thames, S.F., Kaleem, K., 1991. Guayule resin in amine-epoxy strippable coatings. *Bioresour. Technol.* 35, 185–190. [https://doi.org/10.1016/0960-8524\(91\)90028-I](https://doi.org/10.1016/0960-8524(91)90028-I).
- Thames, S.F., Wagner, J.P., 1991. Recent advances in guayule coprodut research and development. In: Whitworth, W.J., Whitehead, E.E. (Eds.), *Guayule Natural Rubber: a Technical Publication with Emphasis on Recent Findings*. GAMC-USDA/CSRS, Tucson, AZ, pp. 311–350.
- Thames, S., Schuman, T., Reichel, L., Purvis, W., Poole, P., 1996. Guayule Coproduts: Emerging Technology in Industrial, Marine, and Peelable Coatings. *Int SAMPE Symp Exhib.*
- Traub, H.P., Slattery, M.C., 1946. Analysis of levulins, inulin, and monosaccharides in guayule. *Bot. Gaz.* 108, 295–299. <https://doi.org/10.1086/335416>.
- van Beilen, J.B., Poirier, Y., 2007. Guayule and Russian dandelion as alternative sources of natural rubber. *Crit. Rev. Biotechnol.* 27, 217–231. <https://doi.org/10.1080/07388550701775927>.
- Wagner, J.P., Parma, D.G., 1988. Continuous solvent extraction process for recovery of natural rubber from guayule. *Polym. Technol. Eng.* 27, 335–350. <https://doi.org/10.1080/03602558808070113>.
- Wagner, J.P., Parma, D.G., 1989. Establishing a domestic guayule natural rubber industry. *Polym. Technol. Eng.* 28, 753–777. <https://doi.org/10.1080/03602558908049825>.
- Wagner, J.P., Schloman Jr., W.W., 1991. Processing. In: Whitworth, W.J., Whitehead, E. E. (Eds.), *Guayule Natural Rubber: a Technical Publication with Emphasis on Recent Findings*. GAMC-USDA/CSRS, Tucson, AZ, pp. 261–286.
- Walter, E.D., 1944. Isolation of partheniol, parthenyl cinnamate, and other constituents from guayule resin I. *J. Am. Chem. Soc.* 66, 419–421. <https://doi.org/10.1021/ja01231a034>.
- Watkins, S.F., Fronczek, F.R., Chiari, G., Reynolds, G.W., Rodriguez, E., 1985. Molecular structure of Guayulin a. *J. Nat. Prod.* 48, 631–633. <https://doi.org/10.1021/np50040a018>.
- West, J., Rodriguez, E., Hashemi, A., 1991. Biochemical evolution and species relationships in the genus *Parthenium* (Asteraceae). In: Whitworth, W.J., Whitehead, E.E. (Eds.), *Guayule Natural Rubber: a Technical Publication with Emphasis on Recent Findings*. GAMC-USDA/CSRS, Tucson, AZ, pp. 17–31.

- Wink, M., 2010. Annual Plant Reviews, Volume 40, Biochemistry of Plant Secondary Metabolism, 2nd ed. Blackwell Publishing, p. 464.
- Wright, G.N., Fansler, S., Lacewell, R.D., 1991. Guayule economics. In: Whitworth, J.W., Whitehead, E.E. (Eds.), Guayule Natural Rubber: a Technical Publication with Emphasis on Recent Findings. GAMC-USDA/CSRS, Tucson, AZ, pp. 351–365.
- Yarnell, E., 2007. Plant chemistry in veterinary medicine: medicinal constituents and their mechanisms of action. CHAPTER 11. In: Wynn, S.G., Fougère, B.J. (Eds.), Veterinary Herbal Medicine. Mosby, Saint Louis, pp. 159–182.
- Zoeller Jr., J.H., Wagner, J.P., Sulikowski, G.A., 1994. Concise multigram purification of guayulin A from guayule. J. Agric. Food Chem. 42, 1647–1649. <https://doi.org/10.1021/jf00044a012>.